



Skyline

• Tutorial Webinar  
#17

# PRM Method Development and Data Analysis with Skyline

With

Brendan MacLean (Principal Developer, Skyline)  
Eduard Sabidó, Ph.D. (Head of the UPF/CRG Proteomics Unit)  
Cristina Chiva, Ph.D. (PRM researcher, CRG Proteomics Unit)

# Agenda

- Welcome from the Skyline team!
- PRM Method Development and Data Analysis with Skyline
  - Introduction with Brendan MacLean
  - Theoretical concepts and benefits of PRM with Eduard Sabidó
  - Tutorial with Cristina Chiva
- Audience Q&A – submit questions to Google Form:

<https://skyline.ms/QA4Skyline.url>

# It Began as Targeted MS/MS (pseudo-SRM)

- ASMS 2011 – Poster presentation (Birgit Schilling)
  - *Skyline: Targeted Proteomics with Extracted Ion Chromatograms from Full-Scan Mass Spectra*
- JPR May, 2012 – Sherrod, et. al (started by Amy Ham, 2009)
  - *Label-Free Quantitation of Protein Modifications by Pseudo-Selected Reaction Monitoring with Internal Reference Peptides*
- MCP Nov, 2012 – Peterson, et. al (Coon lab) – named **PRM**
  - *Parallel reaction monitoring for high resolution and high mass accuracy quantitative, targeted proteomics*
- Anal. Chem. 2015 – Schilling, et. al – latest on Skyline PRM for HRMS
  - *Multiplexed, Scheduled, High-Resolution Parallel Reaction Monitoring on a Full Scan QqTOF...*

# Recorded Webinars

- 16 other webinars + this one *Coming Soon!*
- **Webinar #3 (2013)**  
PRM Targeted Proteomics Using Full-Scan MS and Skyline  
**Bruno Domon**
- **Webinar #9 (2015)**  
PRM for PTM studies with Skyline  
Research-grade targeted proteomics assay development  
**Jacob D. Jaffe**

# Interest Remains High

- Modern instruments support PRM well
  - High resolution, faster cycle times and full scheduling
- Viable alternative to SRM with triple quadrupole instrument
- Appealing not to need an extra instrument for targeted
  
- Registration for first PRM webinar (431) v 2014 DIA webinar (342)
- Registration for this PRM webinar (434) v 2017 DIA webinar (409)

Eduard Sabidó



**Webinar**

## PRM theoretical concepts, benefits and instrument acquisition settings

Eduard Sabidó  
Cristina Chiva

*CRG/UPF Proteomics Unit  
Barcelona, Spain*



Barcelona  
Biomedical  
Research  
Park



Eduard Sabidó



Cristina Chiva







## PRM is a targeted proteomics workflow

*“Targeted proteomics detects proteins of interest with high **sensitivity**, quantitative **accuracy** and **reproducibility**”*

*“By delivering **precise, reproducible** quantification of proteins of interest in biological samples, targeted proteomics approaches are allowing researchers to apply the scientific method using mass spectrometry”*

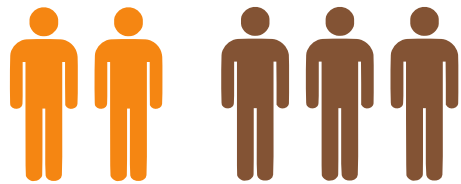
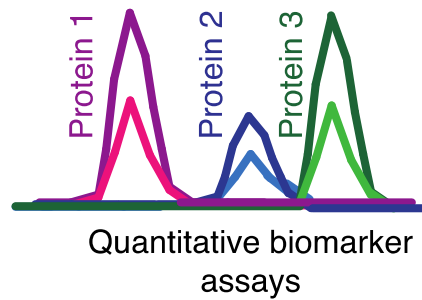




## PRM is a targeted proteomics workflow

### Types of projects suited for targeted proteomics

#### **A** biomarkers



Treatment A

Treatment B

#### **B** pathways

#### **C** interactions





## PRM is a targeted proteomics workflow

### MS1 Targeted Methods

*They rely on the  
mass of the entire molecule*

### MS2 Targeted Methods

*They rely on the  
fragments of the molecule*

### Targeted Acquisition

*They only acquire the  
molecules of interest*

### Targeted Data Analysis

*They acquire everything and later  
specific information is extracted*

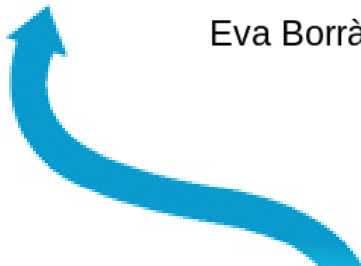
Viewpoint

**What is targeted proteomics? A concise revision of targeted acquisition and targeted data analysis in mass spectrometry**



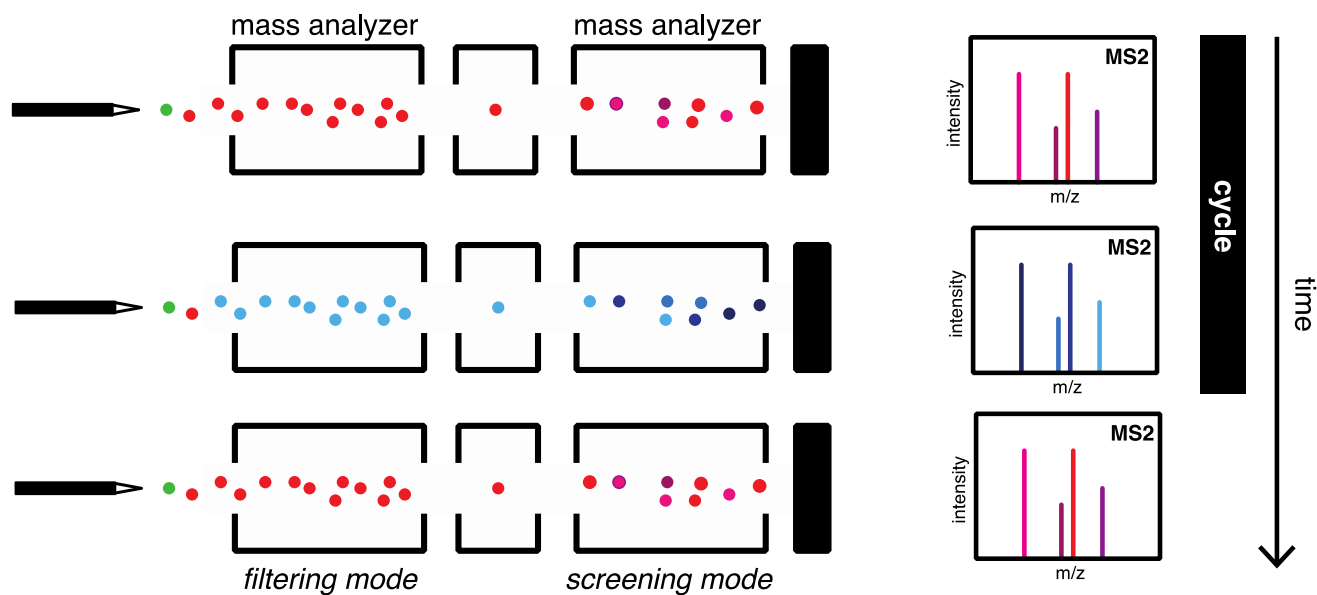
Eva Borràs<sup>1,2</sup> and Eduard Sabidó<sup>1,2,\*</sup>

Issue — *Proteomics* 17, 17–18, 2017, 1700180



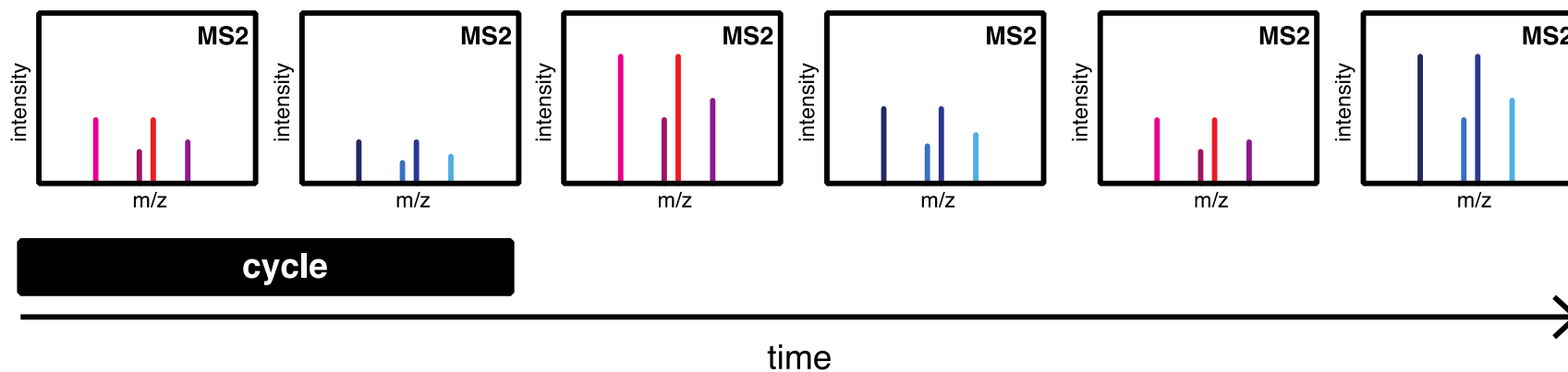
## PRM is a targeted proteomics workflow

### *Parallel Reaction Monitoring (PRM)*



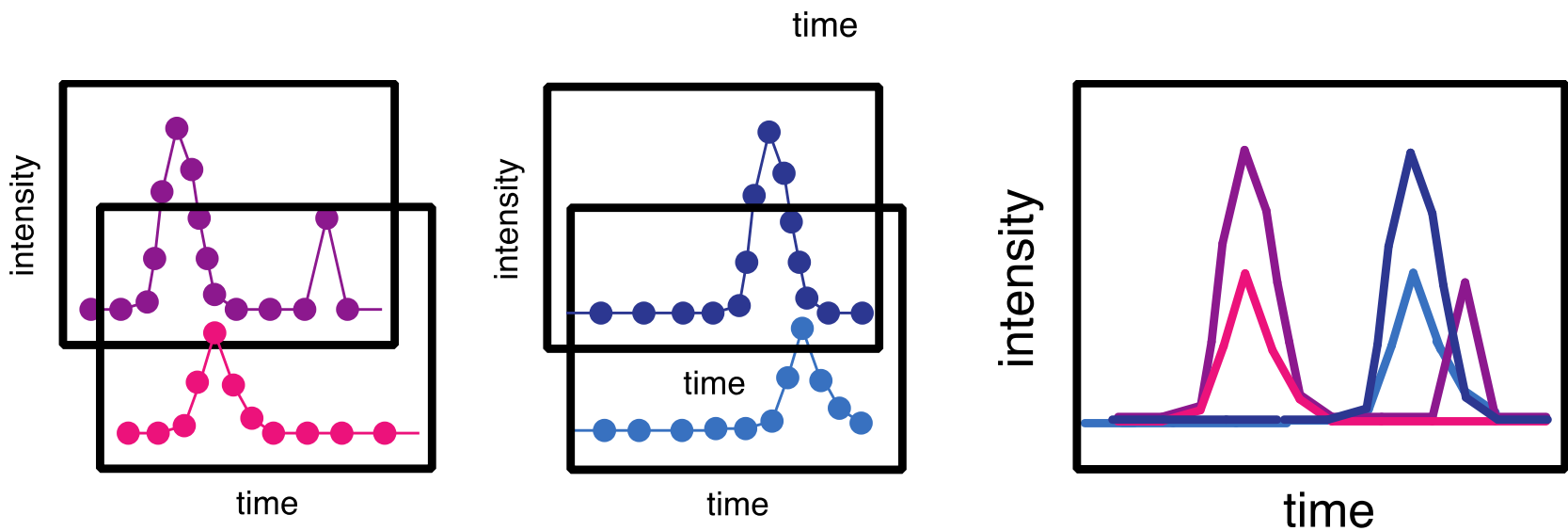
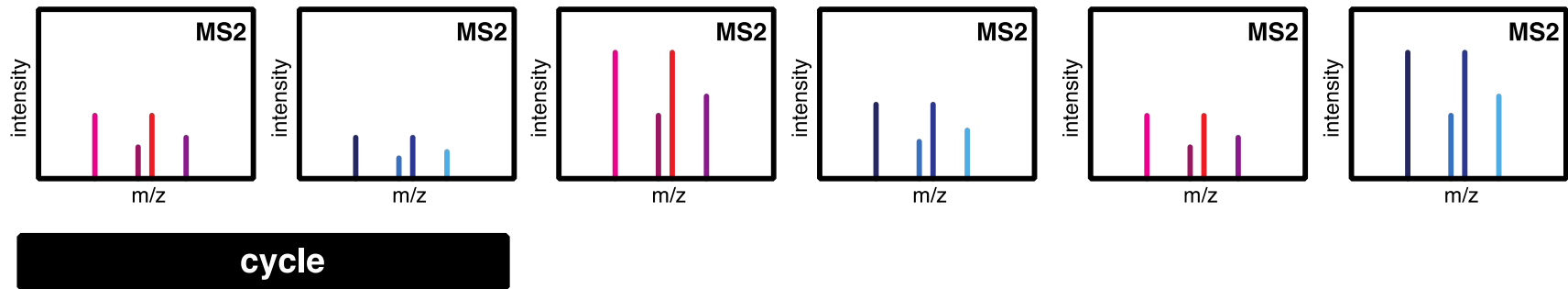
## PRM is a targeted proteomics workflow

### *Parallel Reaction Monitoring (PRM)*



## PRM is a targeted proteomics workflow

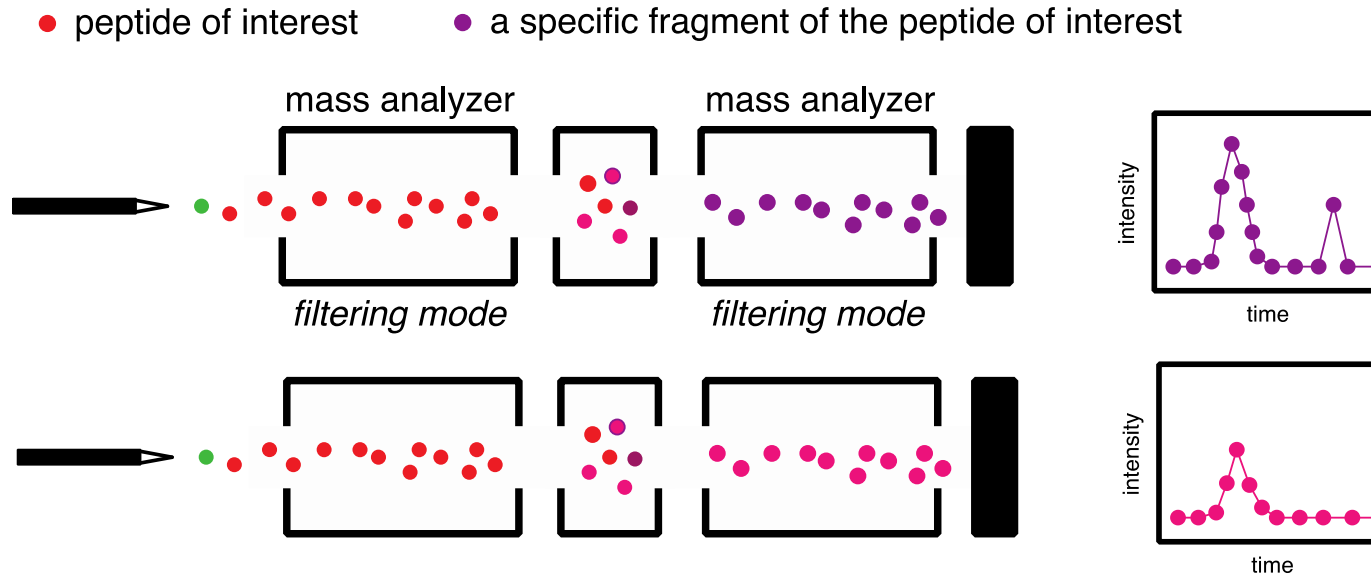
### Parallel Reaction Monitoring (PRM)



co-elution  
identification

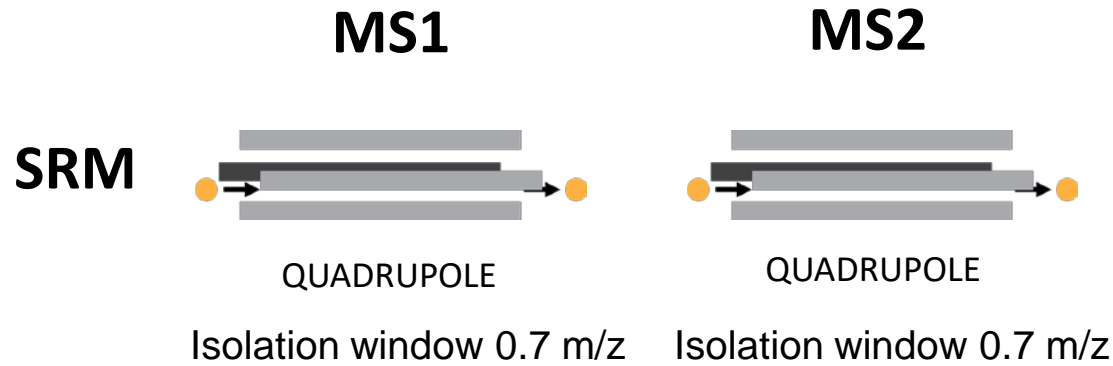
## PRM is a targeted proteomics workflow

*Selected Reaction Monitoring (SRM) is sequential (not parallel)*

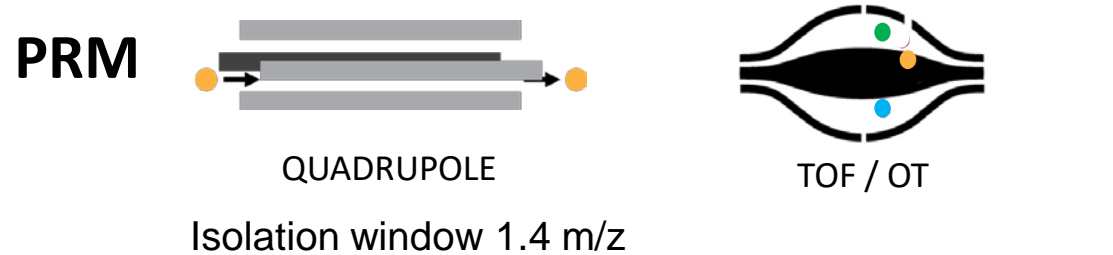
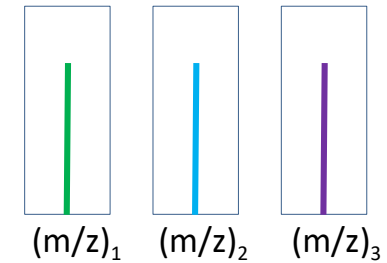


# PRM is a targeted proteomics workflow

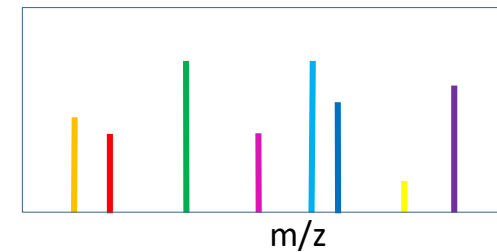
## Mass spectrometry instruments



Low-resolution MS2 signal  
One narrow scan per fragment  
Hardware selected MS2 fragments



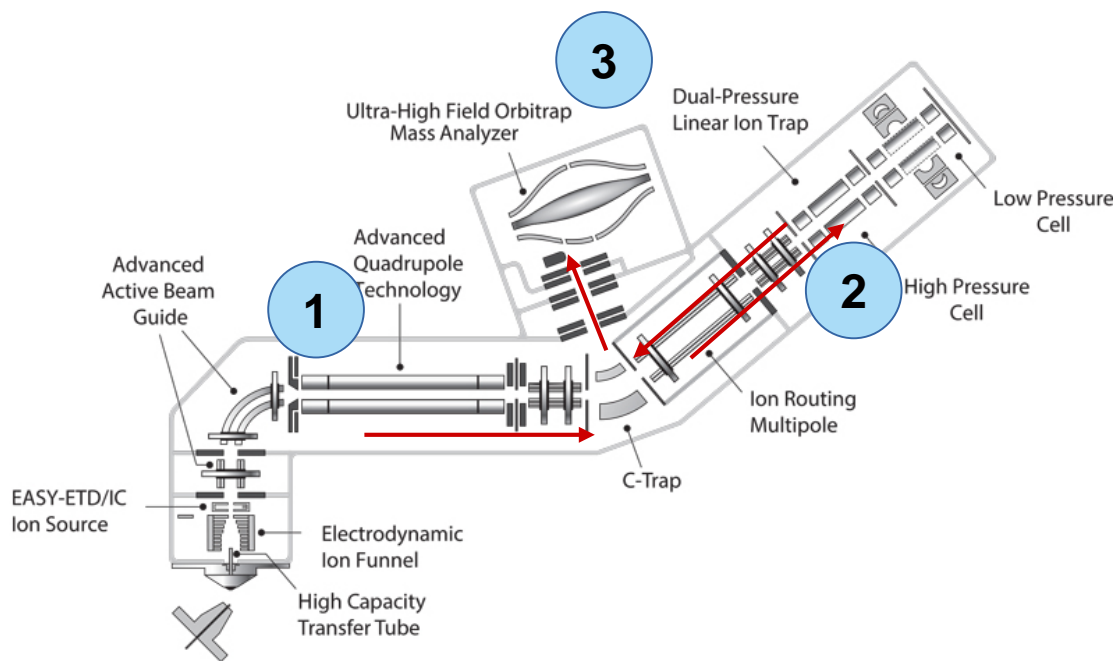
High-resolution MS2 signal  
Full scan MS2  
Software selected MS2 fragments





## PRM is a targeted proteomics workflow

### *PRM in the Orbitrap Fusion Lumos*



1. Precursor selection in quadrupole
  - Isolation window
2. Precursor fragmentation in collision cell
  - Fill time
  - Collision energy
3. Fragment ion detection in the Orbitrap
  - Resolving power



## Peptide verification

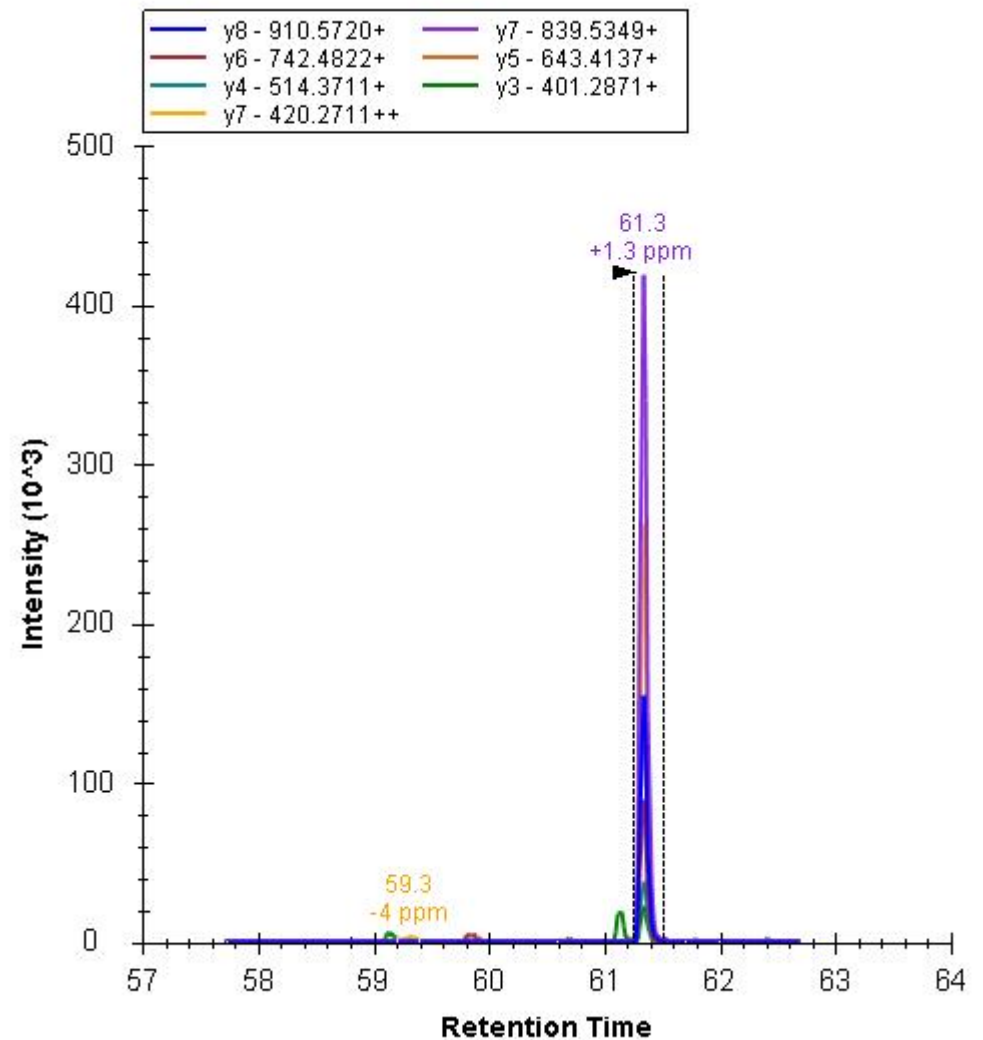
Making sure that the integrated signal corresponds to the targeted peptide



## Peptide verification

Making sure that the integrated signal corresponds to the targeted peptide

### 1. Co-elution of concurrent transitions

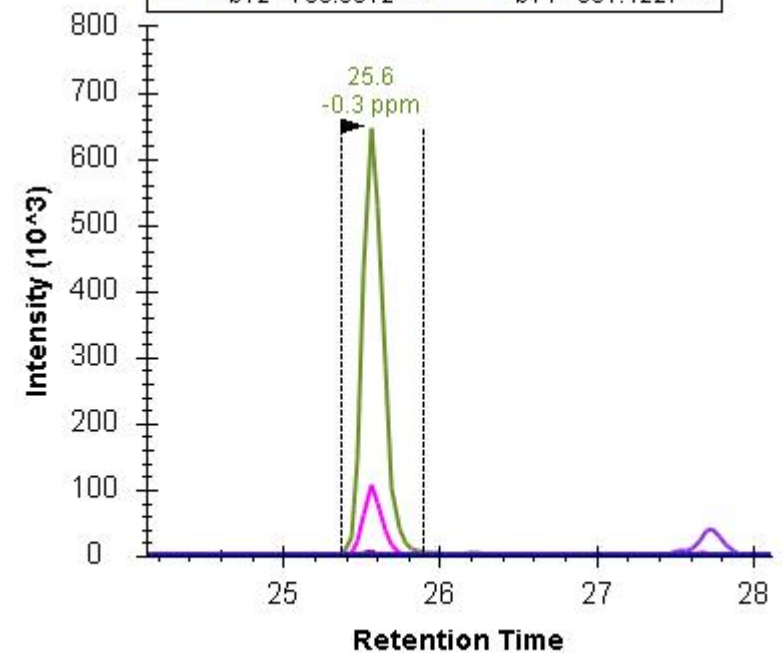


## Peptide verification

Making sure that the integrated signal corresponds to the targeted peptide

1. Co-elution of concurrent transitions
2. Sequence information / coverage

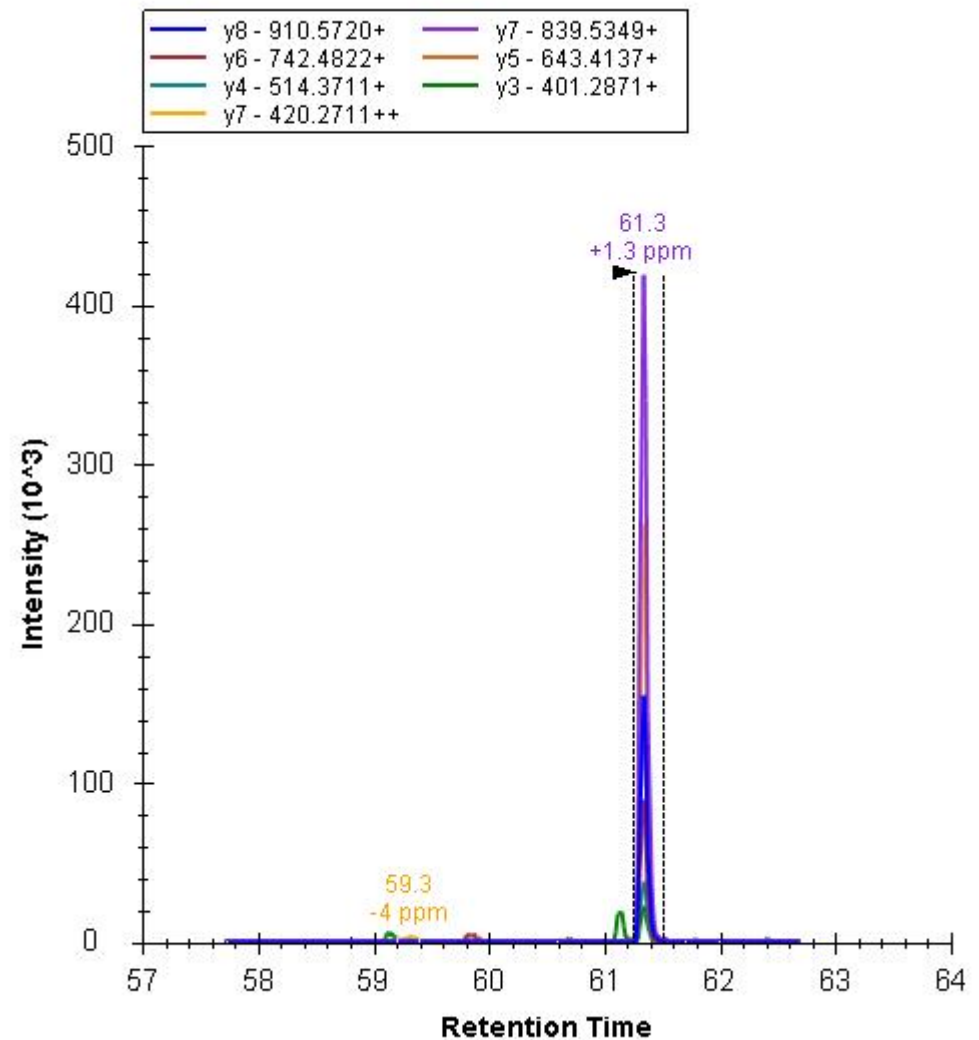
y9 - 1110.5790+	y8 - 997.4949+
y7 - 868.4523+	y6 - 740.3937+
y5 - 625.3668+	y4 - 512.2827+
y3 - 397.2558+	y14 - 847.9334++
y13 - 783.4121++	y12 - 726.3907++
y11 - 669.8486++	y10 - 620.3144++
y9 - 555.7931++	y8 - 499.2511++
y7 - 434.7298++	y6 - 370.7005++
b3 - 357.1769+	b4 - 470.2609+
b5 - 569.3293+	b6 - 698.3719+
b7 - 811.4560+	b8 - 940.4986+
b9 - 1068.5572+	b10 - 1183.5841+
b6 - 349.6896++	b7 - 406.2316++
b8 - 470.7529++	b10 - 592.2957++
b12 - 706.3512++	b14 - 831.4227++



## Peptide verification

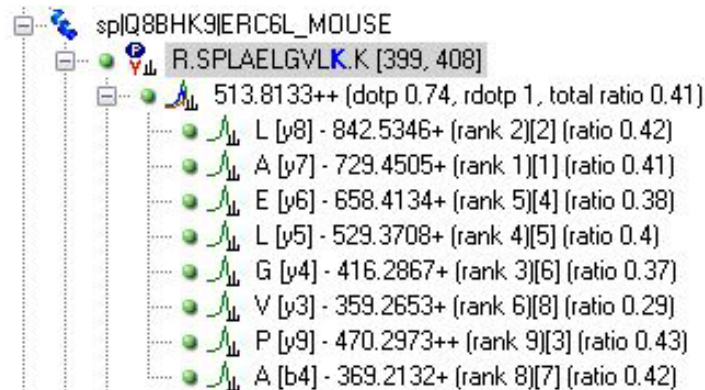
Making sure that the integrated signal corresponds to the targeted peptide

1. Co-elution of concurrent transitions
2. Sequence information / coverage
3. Reference MS2 spectra (libraries)

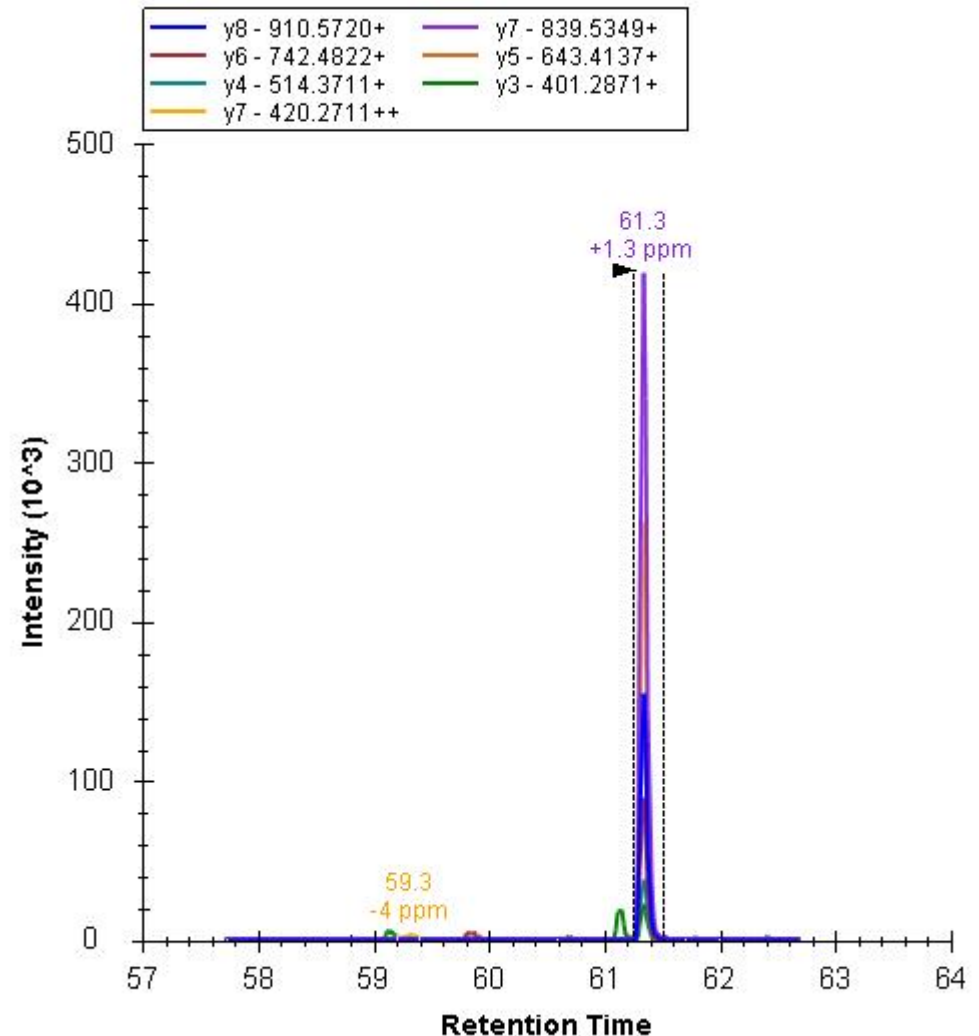
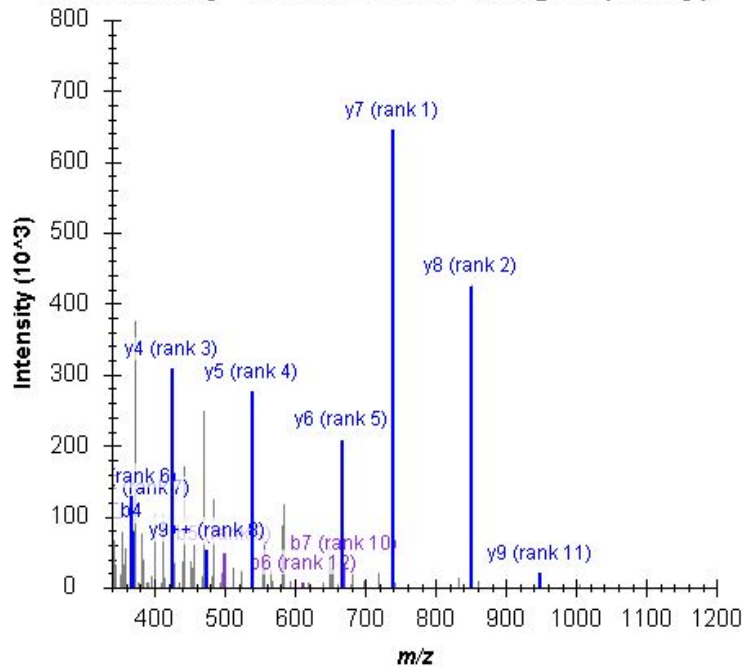


## Peptide verification

Making sure that the integrated signal corresponds to the targeted peptide



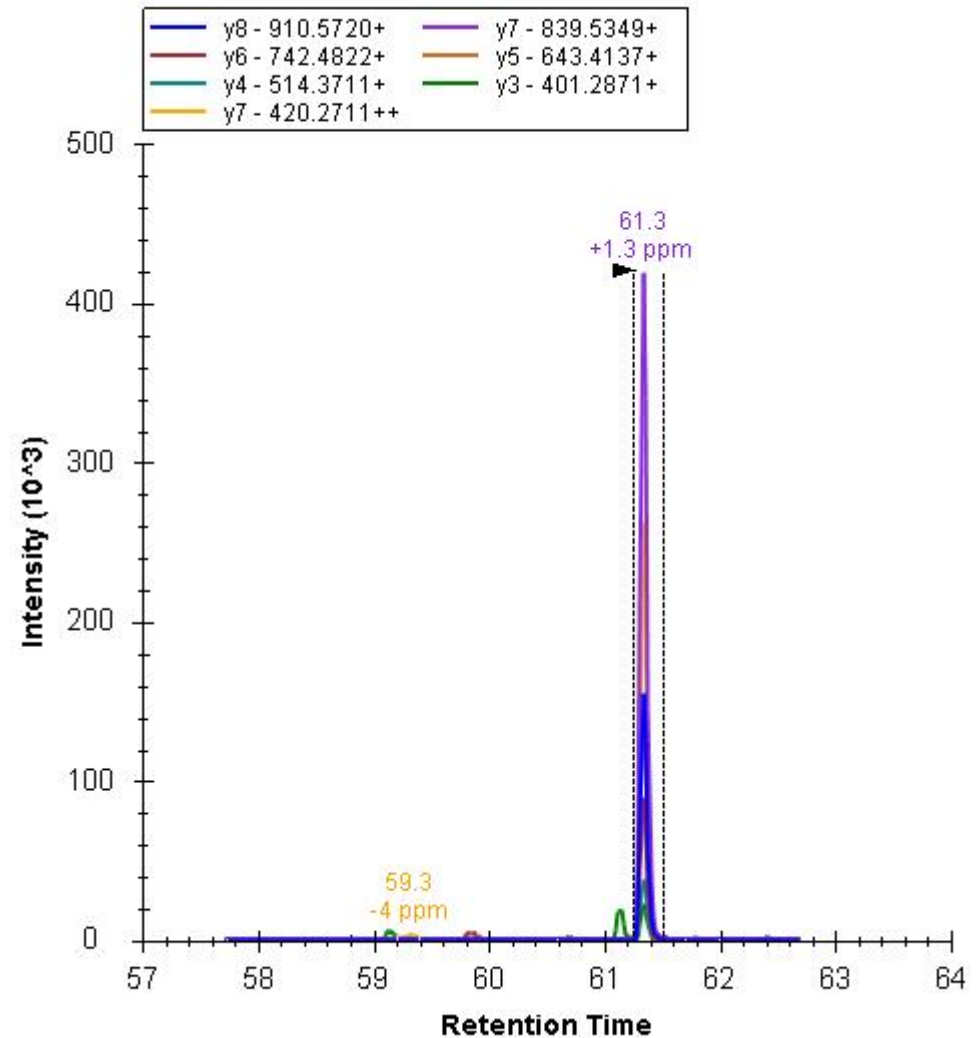
### embo-heavy - SPLAELGVLK, Charge 2 (heavy)



## Peptide verification

Making sure that the integrated signal corresponds to the targeted peptide

1. Co-elution of concurrent transitions
2. Sequence information / coverage
3. Reference MS2 spectra (libraries)
4. Retention time information

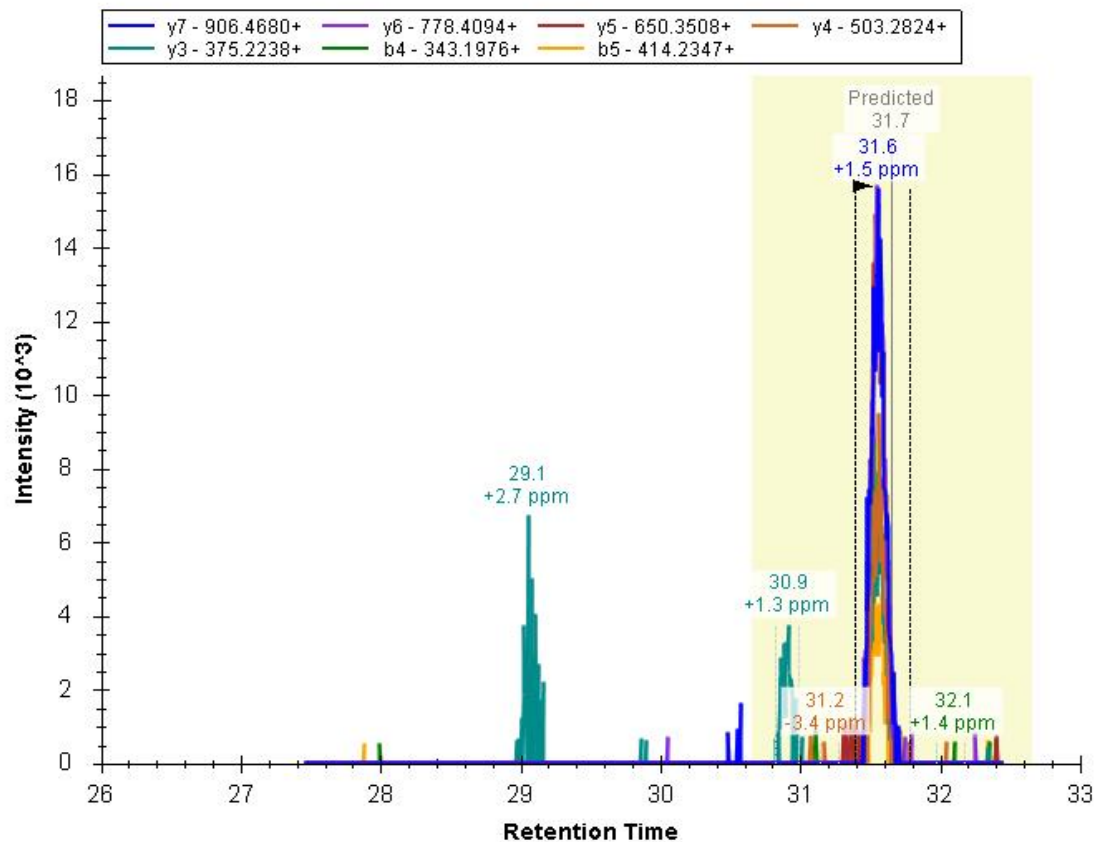
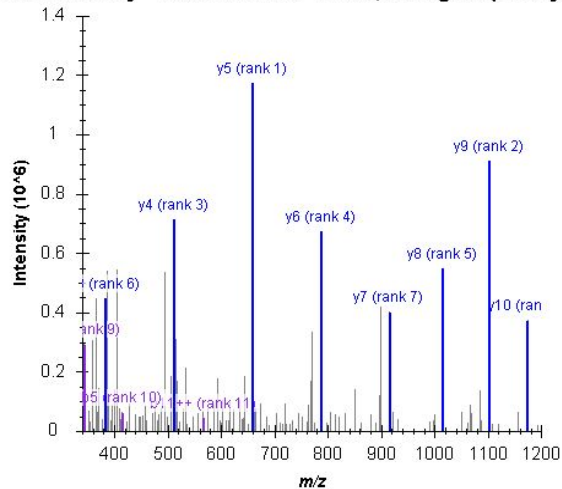


## Peptide verification

Making sure that the integrated signal corresponds to the targeted peptide



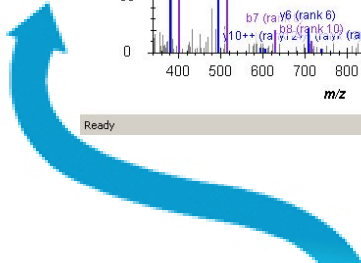
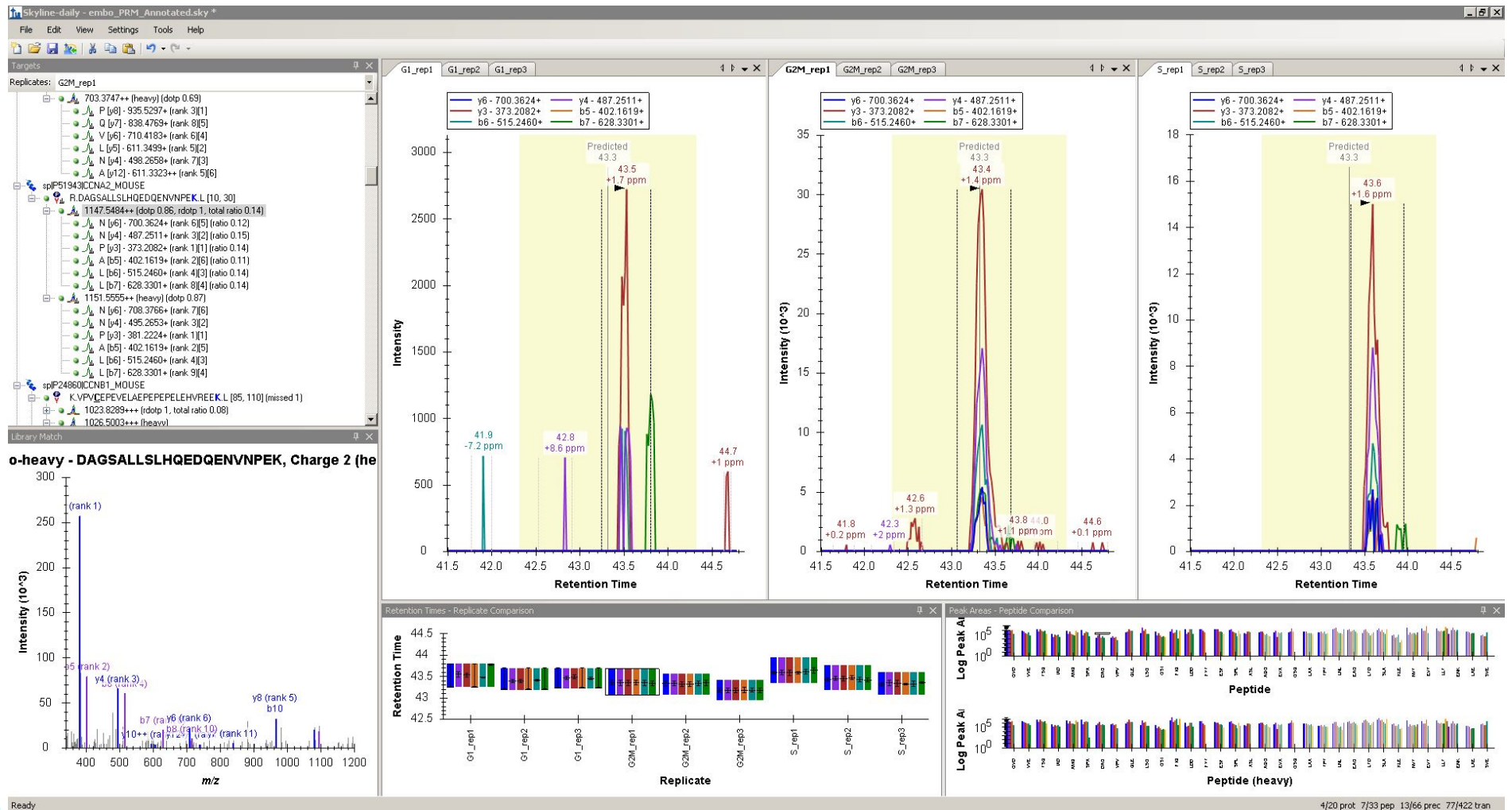
embo-heavy - LAASASTQQFQEVK, Charge 2 (heavy)





# Peptide verification

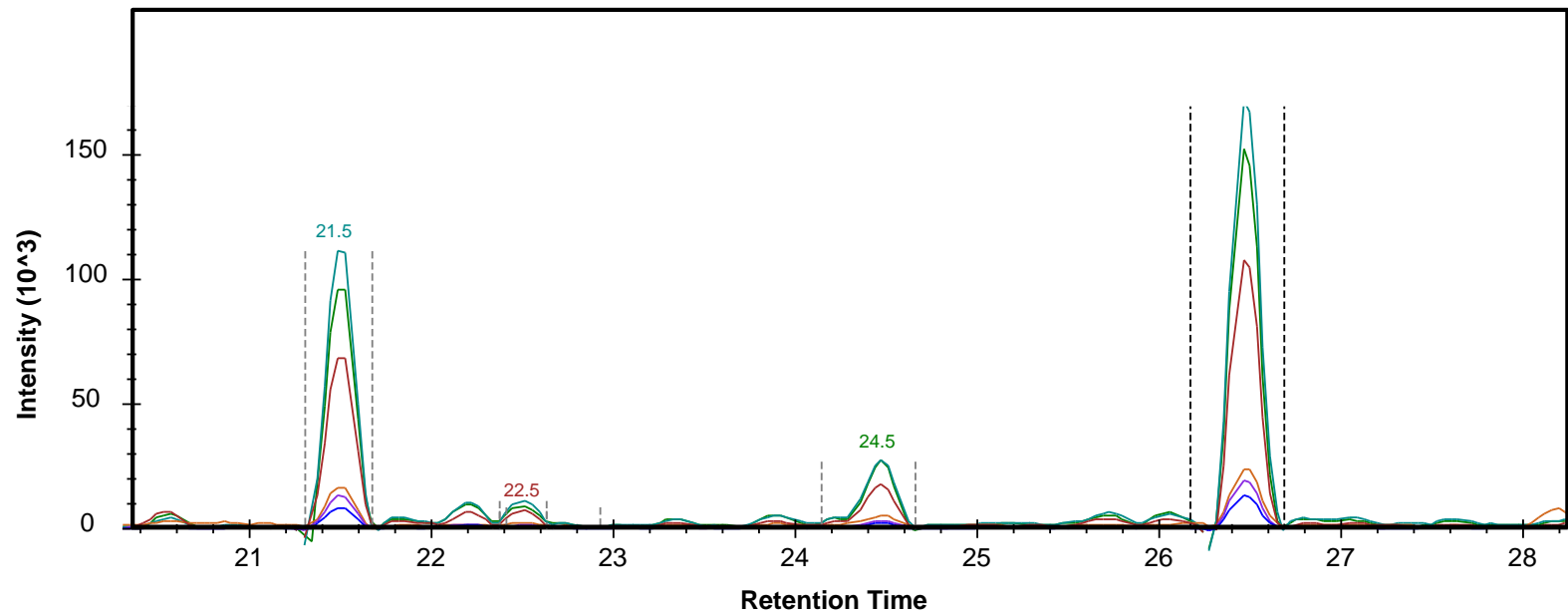
Making sure that the integrated signal corresponds to the targeted peptide





## Peptide verification

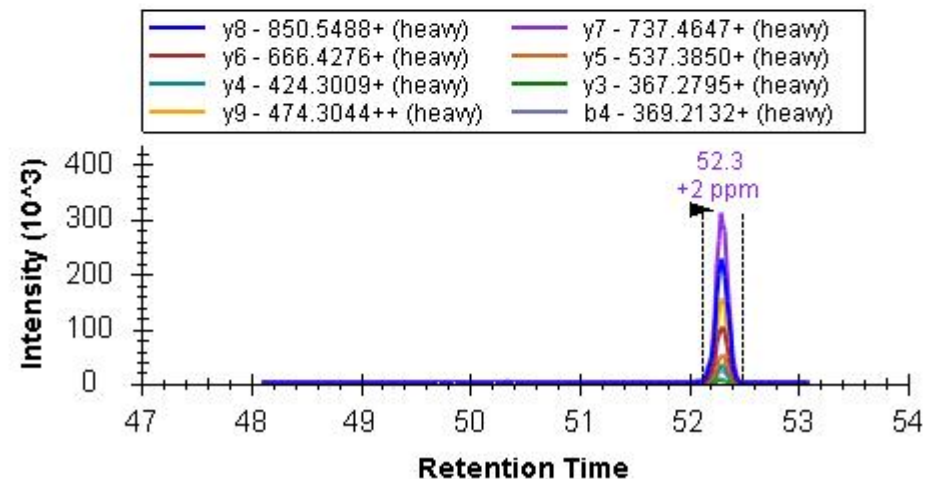
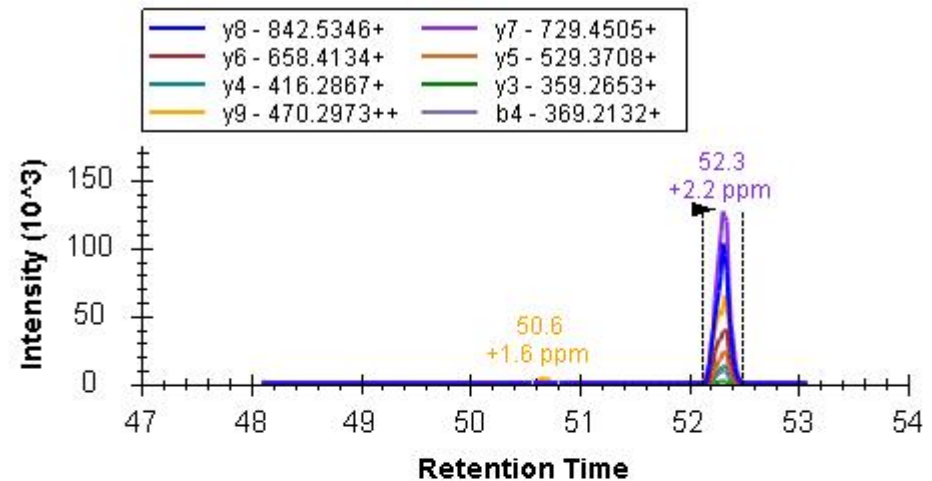
Making sure that the integrated signal corresponds to the targeted peptide



## Peptide verification

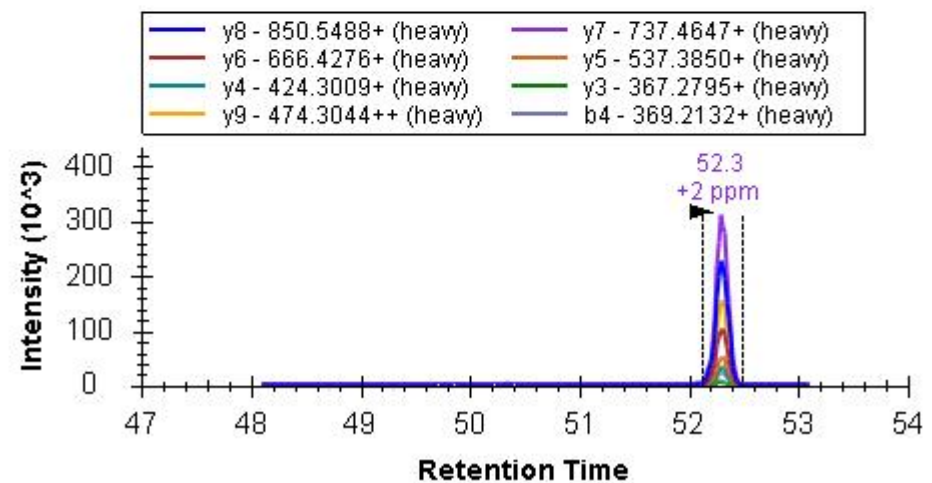
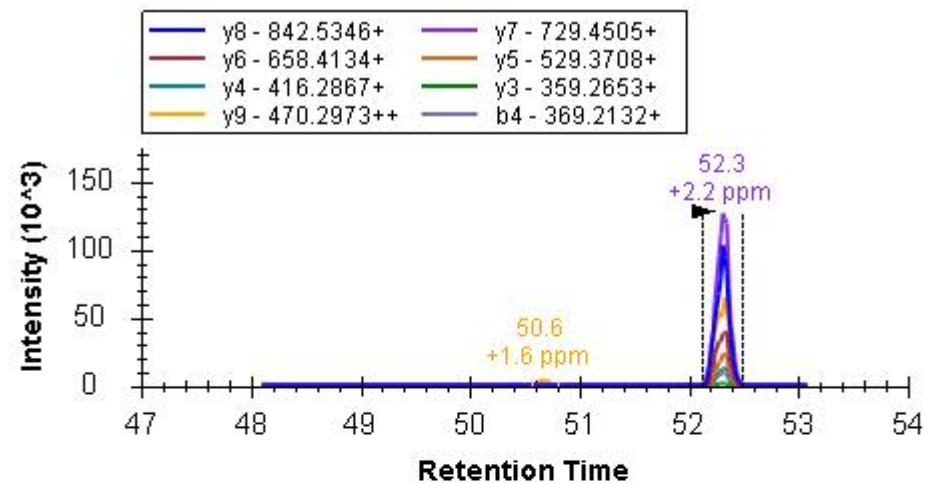
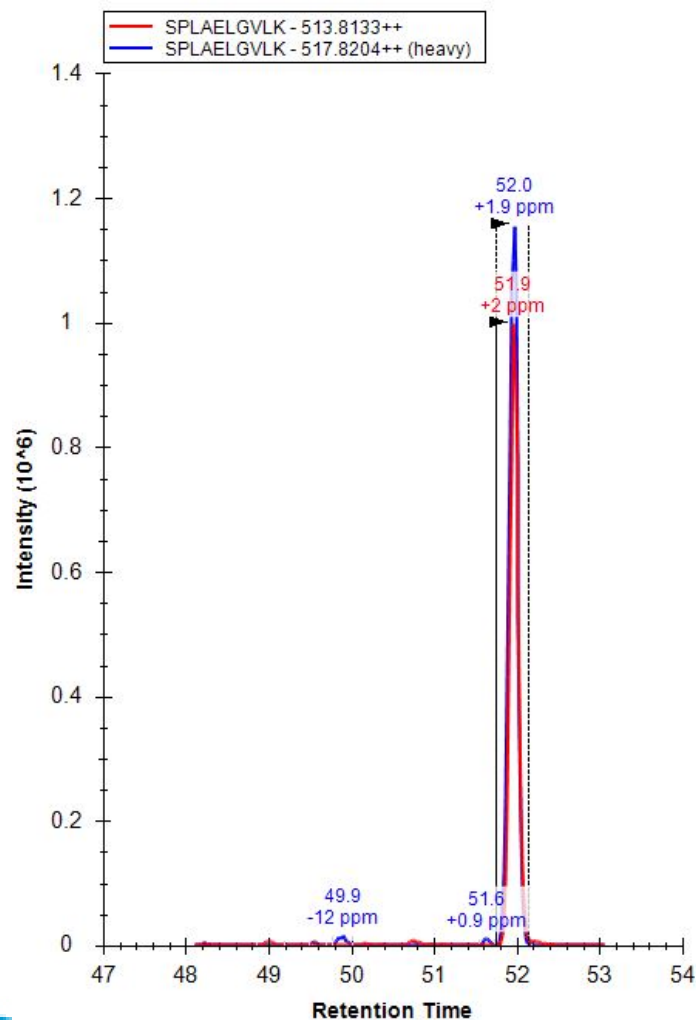
Making sure that the integrated signal corresponds to the targeted peptide

1. Co-elution of concurrent transitions
2. Sequence information / coverage
3. Reference MS2 spectra (libraries)
4. Retention time information
5. Reference internal standard



## Peptide verification

Making sure that the integrated signal corresponds to the targeted peptide





## Peptide quantification

*Aim for a peptide quantification with high accuracy and precision*

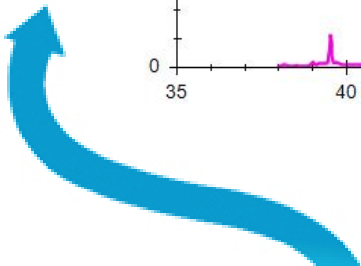
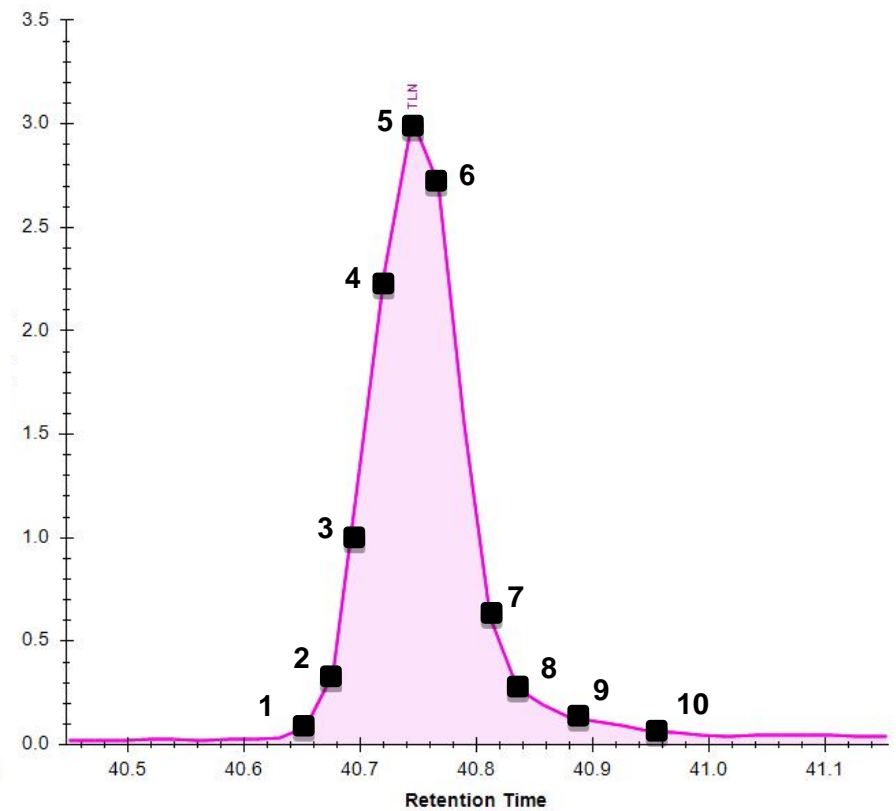
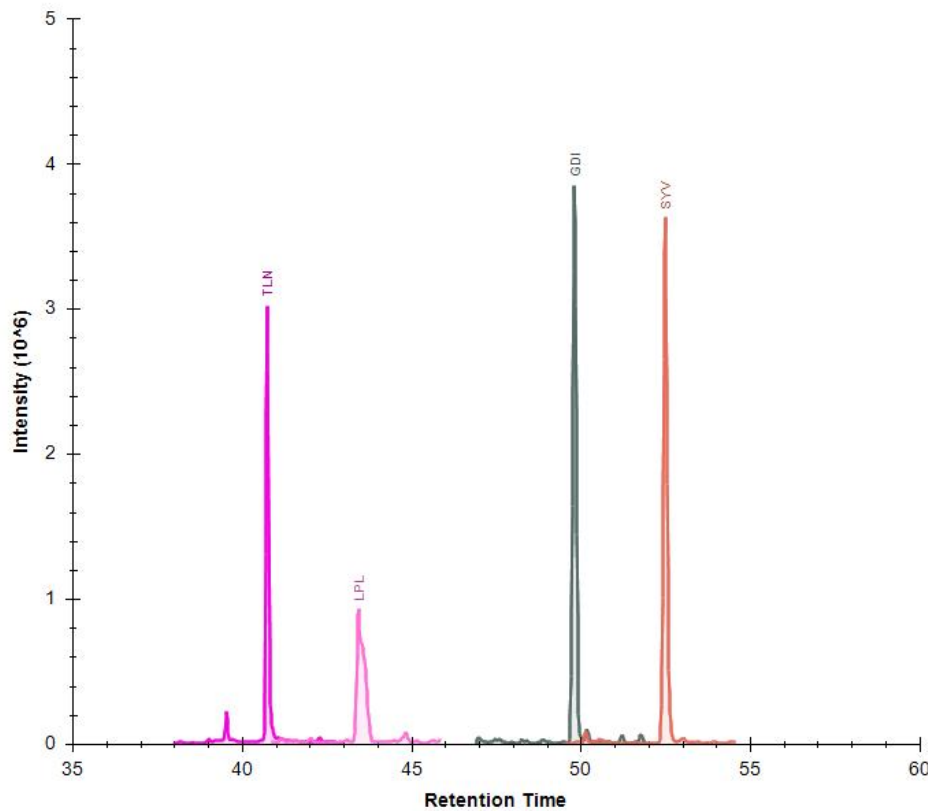




## Peptide quantification

*Aim for a peptide quantification with high accuracy and precision*

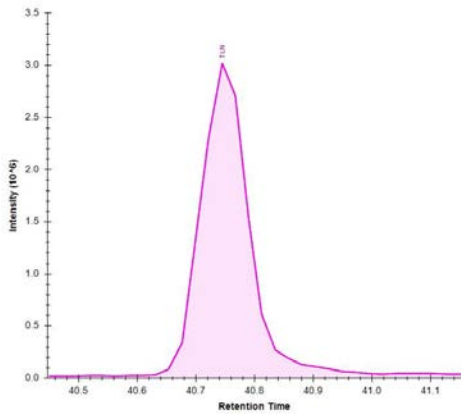
**8-10 points** across chromatographic peak to define elution profile



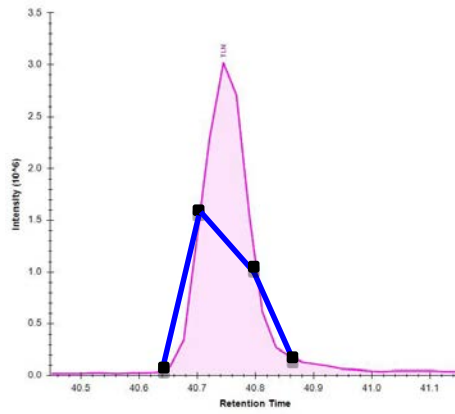
## Peptide quantification

*Aim for a peptide quantification with high accuracy and precision*

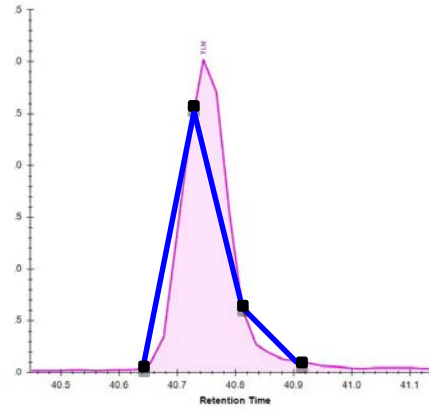
Theoretical



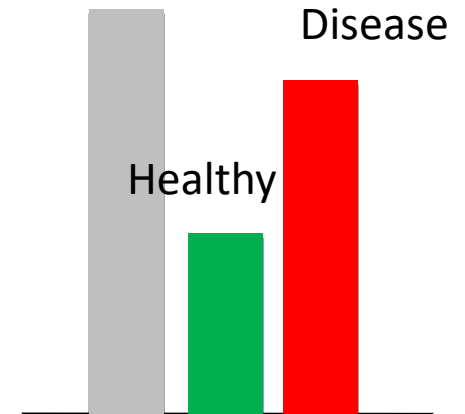
Healthy



Disease



Theoretical



4 points across chromatography peak



## Peptide quantification

*Aim for a peptide quantification with high accuracy and precision*

Calibration strategies with internal standards

1. Single-point calibration
1. External calibration curve
1. Reverse calibration curve

### **Webinar #13 (2016)**

Calibrated Quantification with Skyline  
**Chris Shuford**

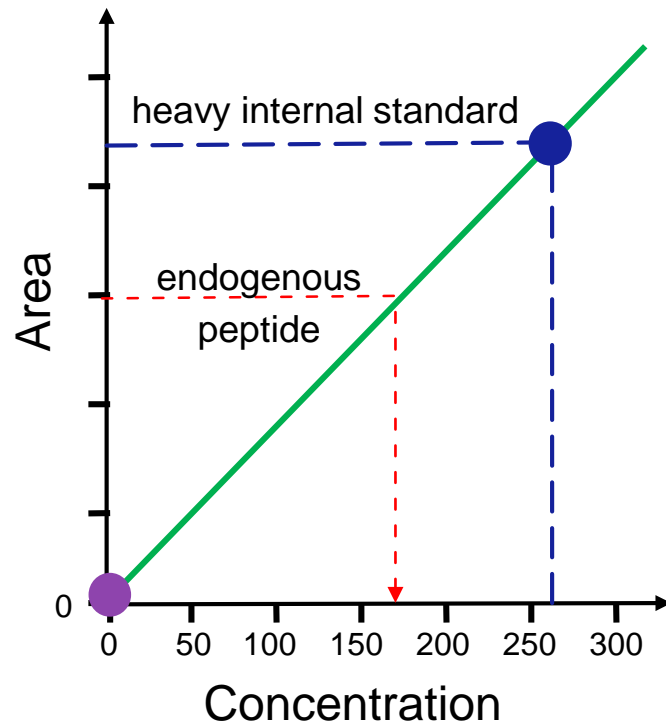




## Peptide quantification

*Aim for a peptide quantification with high accuracy and precision*

### Single-point calibration



We establish the calibration line with two points

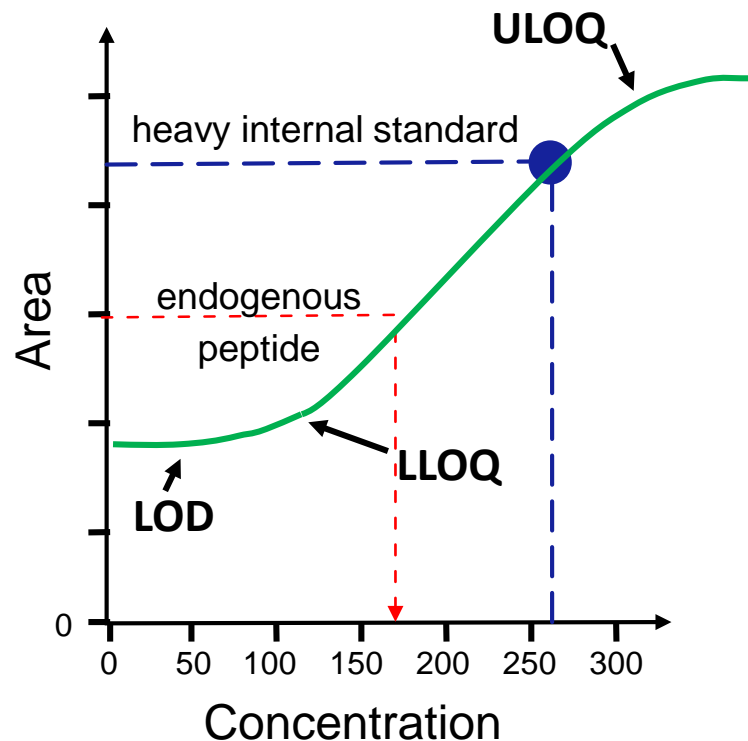
- The response of the heavy internal standard
- The zero — that is, zero response for zero concentration.

We assume a linear range response

## Peptide quantification

*Aim for a peptide quantification with high accuracy and precision*

### Single-point calibration



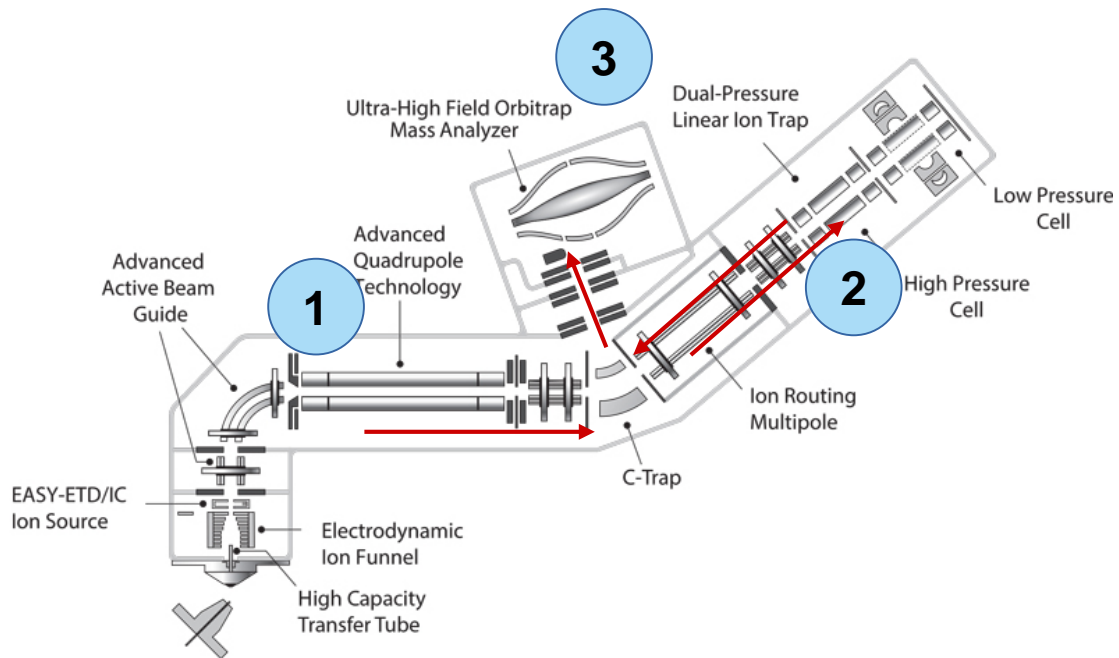
We establish the calibration line with two points

- The response of the heavy internal standard
- The zero — that is, zero response for zero concentration.

We assume a linear range response

## Sensitivity and selectivity: a double boost for PRM

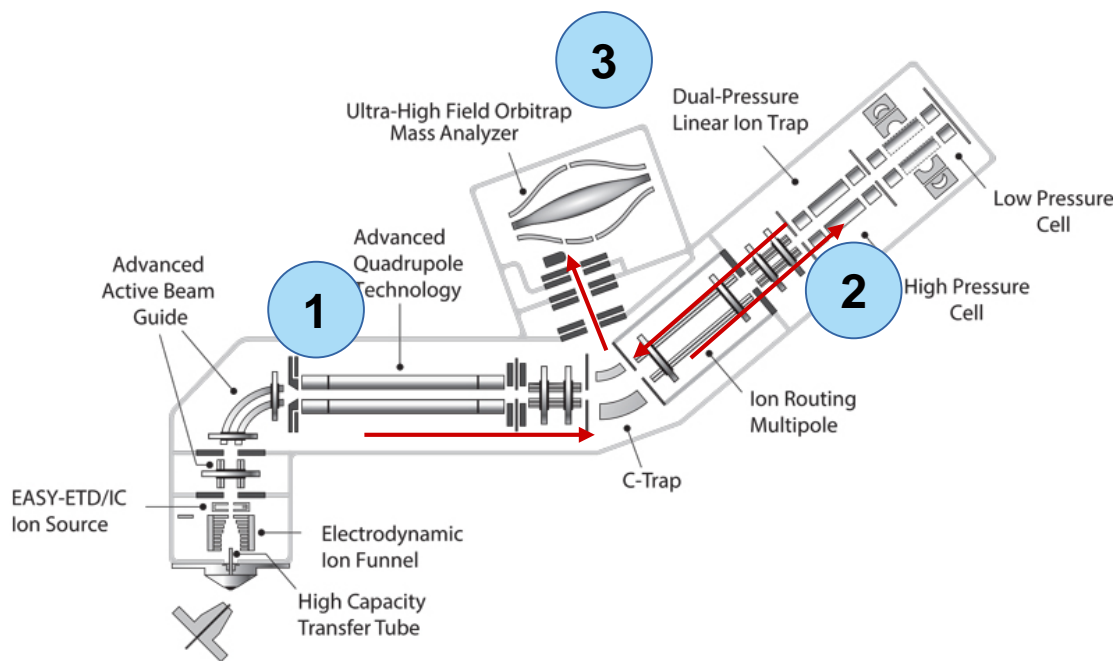
### *PRM in the Orbitrap Fusion Lumos*



1. Precursor selection in quadrupole
  - Isolation window
2. Precursor fragmentation in collision cell
  - Fill time
  - Collision energy
3. Fragment ion detection in the Orbitrap
  - Resolving power

## Sensitivity and selectivity: a double boost for PRM

### *PRM in the Orbitrap Fusion Lumos*



#### 1. Precursor selection in quadrupole

- Isolation window

#### 2. Precursor fragmentation in collision cell

→ SENSITIVITY

- **Fill time**
- Collision energy

#### 3. Fragment ion detection in the Orbitrap

→ SELECTIVITY

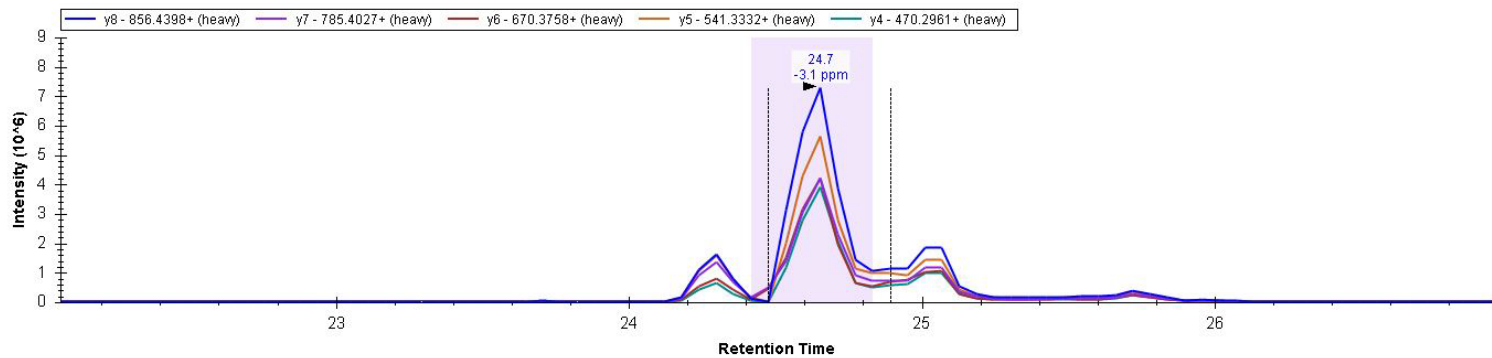
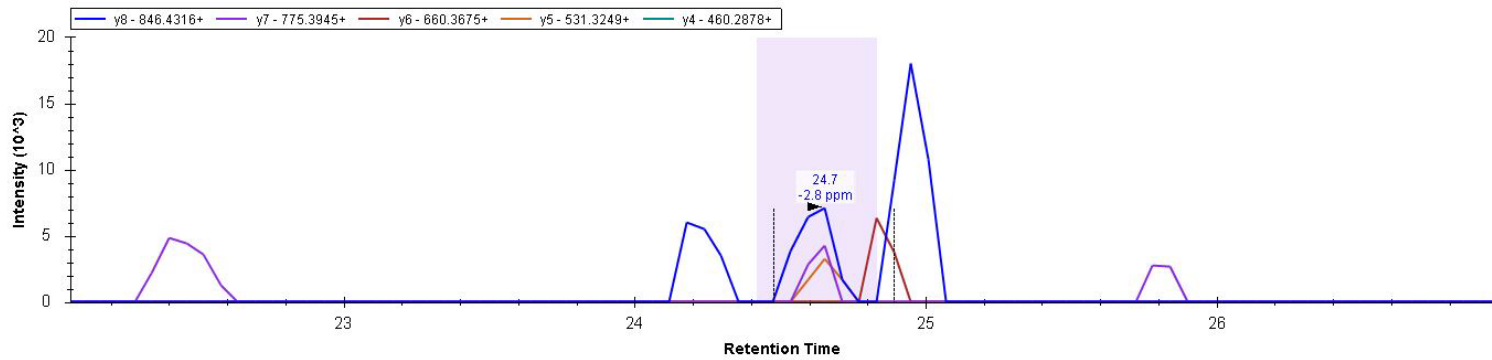
- **Resolving power**

# Sensitivity and selectivity: a double boost for PRM

## *PRM in the Orbitrap Fusion Lumos*

Fill time = 60 ms

Res Power = 30K

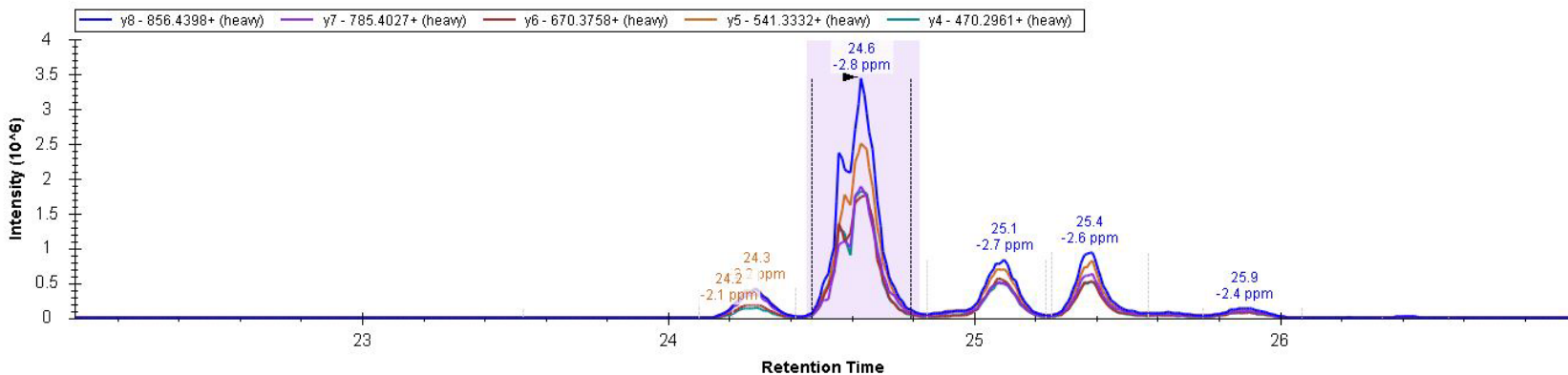
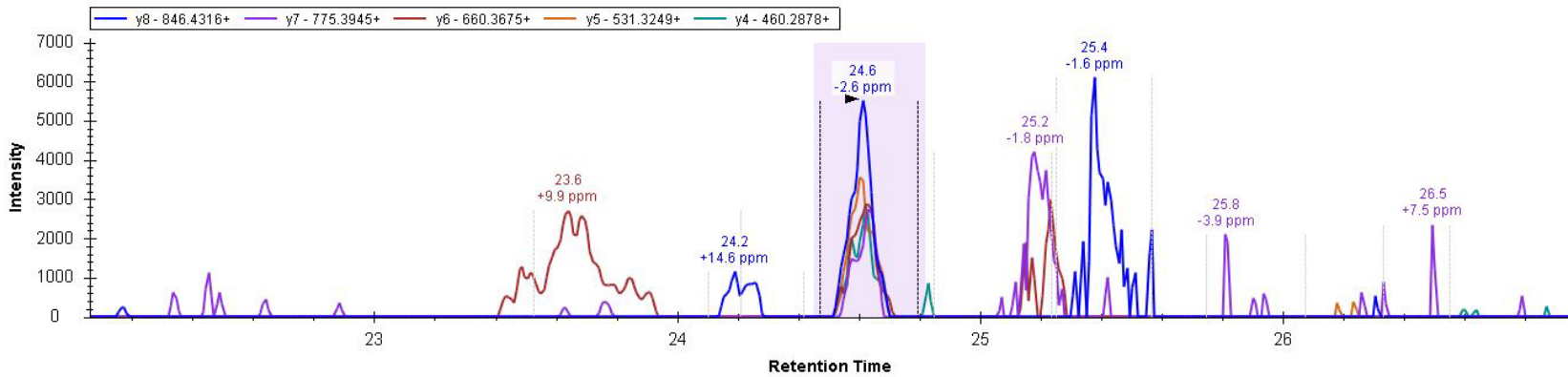


# Sensitivity and selectivity: a double boost for PRM

## PRM in the Orbitrap Fusion Lumos

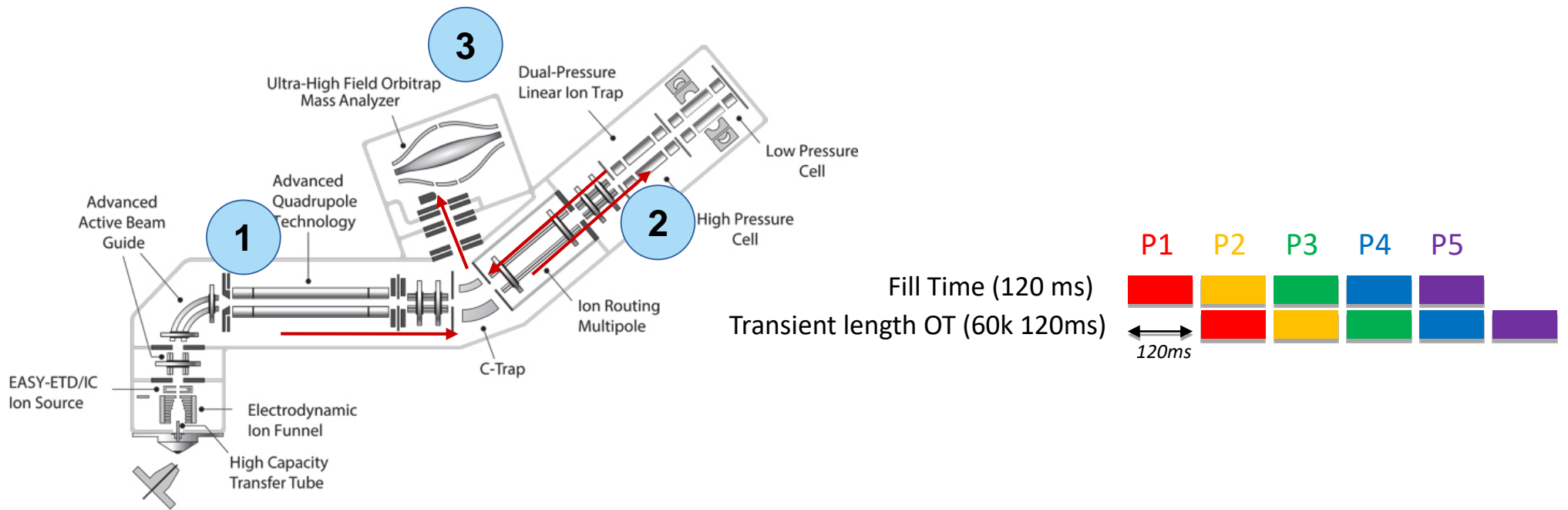
Fill time = 250 ms

Res Power = 120K



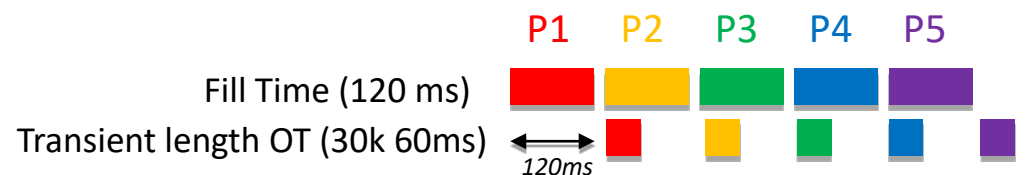
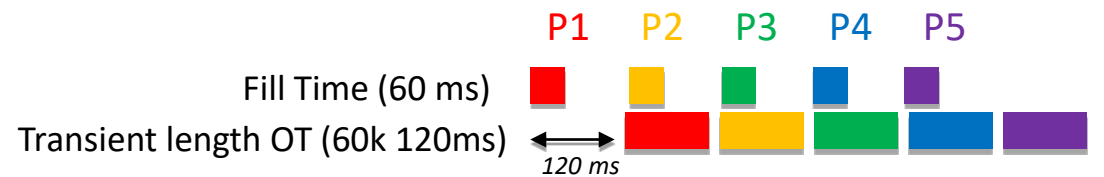
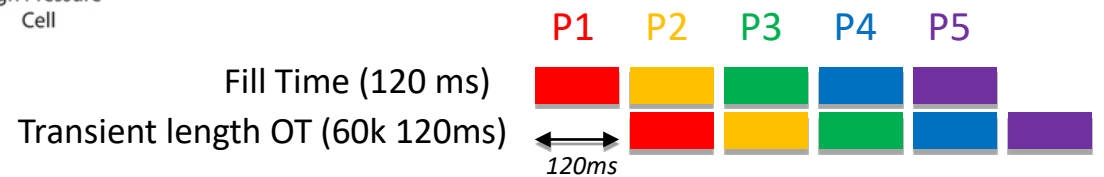
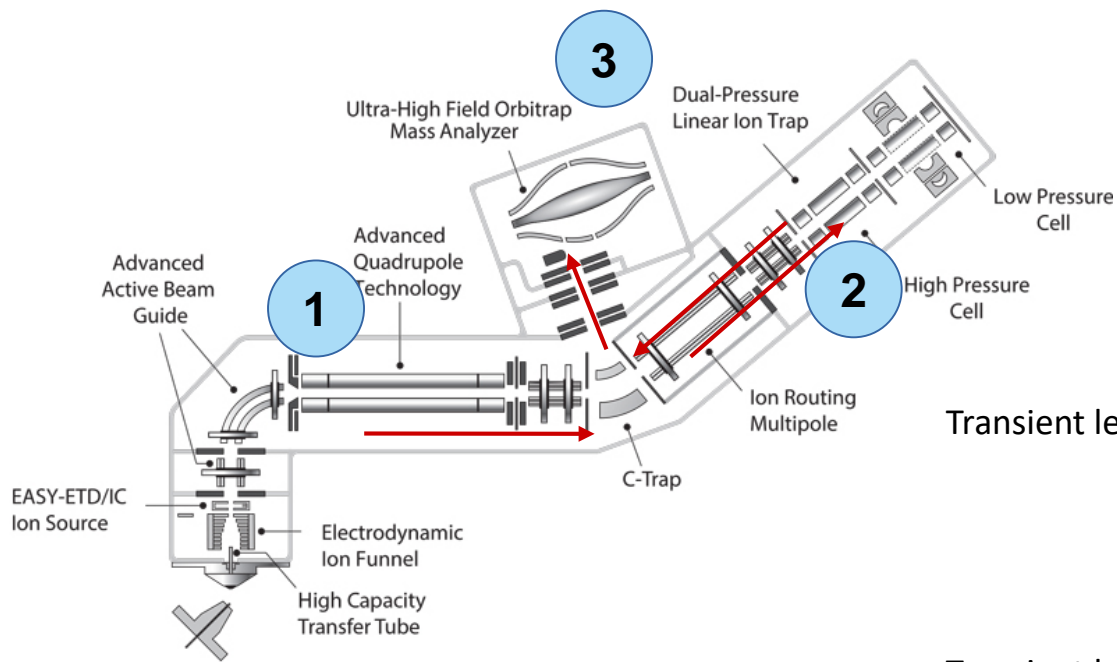
# Sensitivity and selectivity: a double boost for PRM

## PRM in the Orbitrap Fusion Lumos



# Sensitivity and selectivity: a double boost for PRM

## PRM in the Orbitrap Fusion Lumos







## Sensitivity and selectivity: a double boost for PRM

### *PRM in the Orbitrap Fusion Lumos*

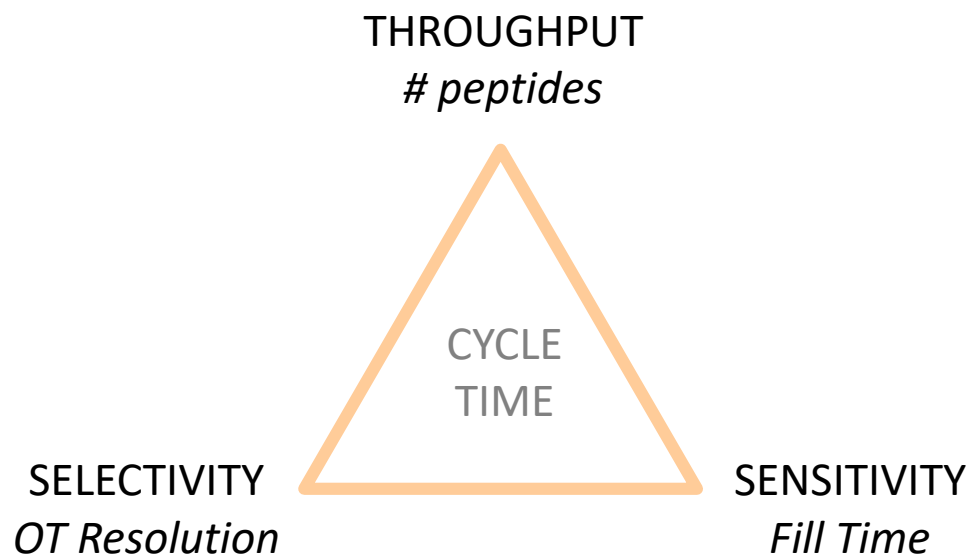
Res. at m/z 200	Transient length [ms]	Approx. scan speed [Hz]	"Free" fill time [ms]
15,000	32	na	22
30,000	64	15	54
60,000	128	7.5	118
120,000	256	4	246
240,000	512	2	502
450,000	1024	<1	1014





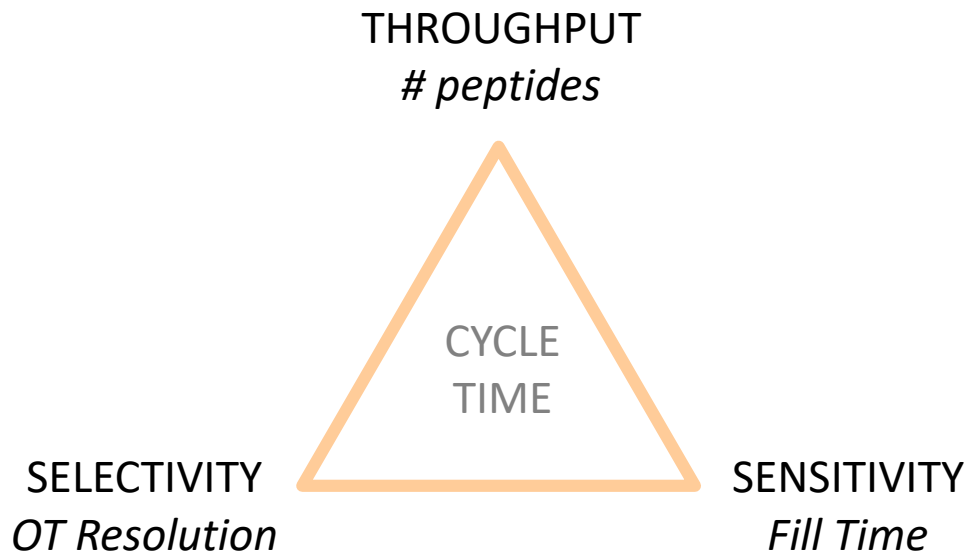
## Sensitivity and selectivity: a double boost for PRM

### *PRM in the Orbitrap Fusion Lumos*

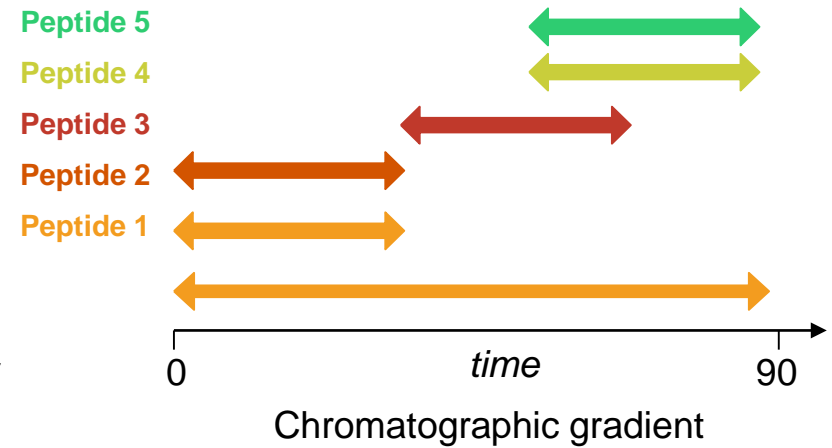


## Sensitivity and selectivity: a double boost for PRM

### *PRM in the Orbitrap Fusion Lumos*



### Scheduled PRM



# Orbitrap Fusion Lumos Method Editor for PRM

Method Editor | Global Parameters | Scan Parameters | Summary

Method Timeline

Method Duration (min): 110

Experiment 1 | Time Range: 0-110 min

Scans

- MS
- MS<sup>n</sup>

Filters

- Precursor Selection Range
- MIPS
- Intensity
- Charge State
- Dynamic Exclusion
- Targeted Inclusion
- Targeted Exclusion
- Apex Detection

Triggers

- Targeted Mass Trigger
- Targeted Loss Trigger

Alternate Precursor Sorts

- Sort by Charge

Targeted MS<sup>n</sup> Scan Properties

- MS<sup>n</sup> Level (n): 2
- Multiplex Ions:
- Isolation Mode: Quadrupole
- Isolation Window (m/z): 1.4
- Activation Type: HCD
- HCD Collision Energy (%): 30
- Stepped Collision Energy:
- Detector Type: Orbitrap
- Orbitrap Resolution: Defined in Table
- Mass Range: Normal
- Scan Range (m/z): 340-950
- RF Lens (%): 30
- AGC Target: 5.0e4
- Inject Ions for All Available Parallelizable Time:
- Maximum Injection Time (ms): Defined in Table
- Microscans: 1
- Data Type: Centroid
- Polarity: Positive
- Source Fragmentation:

Ready

Targeted MS <sup>n</sup> Scan Properties		Show Favorites
MS <sup>n</sup> Level (n)	2	★
Multiplex Ions	<input type="checkbox"/>	★
Isolation Mode	Quadrupole	★
Isolation Window (m/z)	1.4	★
Activation Type	HCD	★
HCD Collision Energy (%)	30	★
Stepped Collision Energy	<input type="checkbox"/>	★
Detector Type	Orbitrap	★
Orbitrap Resolution	Defined in Table	★
Mass Range	Normal	★
Scan Range (m/z)	340-950	★
RF Lens (%)	30	★
AGC Target	5.0e4	★
Inject Ions for All Available Parallelizable Time	<input type="checkbox"/>	★
Maximum Injection Time (ms)	Defined in Table	★
Microscans	1	★
Data Type	Centroid	★
Polarity	Positive	★
Source Fragmentation	<input type="checkbox"/>	★



## Orbitrap Fusion Lumos Method Editor for PRM

Mass List Table										Import	Export	+	×
	Compound	Formula	Adduct	m/z	z	t start (min)	t stop (min)	Orbitrap Resolution	Maximum Injection Time (ms)				
1	IPGIII AASAVR_light			590.8742	2	71.3	81.3	60000	118				
2	IPGIII AASAVR_heavy			595.8784	2	71.3	81.3	60000	118				
3	FGLTTSR_light			391.2138	2	28.52	38.52	60000	118				
4	FGLTTSR_heavy			396.2179	2	28.52	38.52	60000	118				
5	LAALPNVYEVISK_light			708.9085	2	80.26	90.26	60000	118				
6	LAALPNVYEVISK_heavy			712.9156	2	80.26	90.26	60000	118				
7	ASGQAFELISPR_light			694.8803	2	75.58	85.58	30000	54				
8	ASGQAFELISPR_heavy			699.8844	2	75.58	85.58	30000	54				
9	ESVPEFPLSPPK_light			663.8506	2	68.26	78.26	30000	54				
10	ESVPEFPLSPPK_heavy			667.8577	2	68.26	78.26	30000	54				
11	SHEAEVLK_light			456.7429	2	4.98	14.98	60000	118				
12	SHEAEVLK_heavy			460.75	2	4.98	14.98	60000	118				
13	VADYIPQLAK_light			559.3162	2	56.85	66.85	60000	118				
14	VADYIPQLAK_heavy			563.3233	2	56.85	66.85	60000	118				
15	YAIAVNDLGTEYVHR_light			574.2933	3	59.83	69.83	120000	246				
16	YAIAVNDLGTEYVHR_heavy			577.6294	3	59.83	69.83	120000	246				
17	VLSPEAVR_light			435.7558	2	26.21	36.21	120000	246				
18	VLSPEAVR_heavy			440.7599	2	26.21	36.21	120000	246				
19	VLKPIQLTDPGK_light			436.9344	3	40.72	50.72	60000	118				
20	VLKPIQLTDPGK_heavy			439.6058	3	40.72	50.72	60000	118				



## More PRM Webinars

### **Webinar #3 (2013)**

PRM Targeted Proteomics Using Full-Scan MS and Skyline  
**Bruno Domon**

### **Webinar #9 (2015)**

Research-grade targeted proteomics assay development: PRM for PTM studies with Skyline.  
**Jacob D. Jaffe**

### **Other webinars**

<https://skyline.ms/wiki/home/software/Skyline/page.view?name=webinars>





## Hands-on tutorial

*PRM method development and step-by-step analysis of Lumos PRM data*



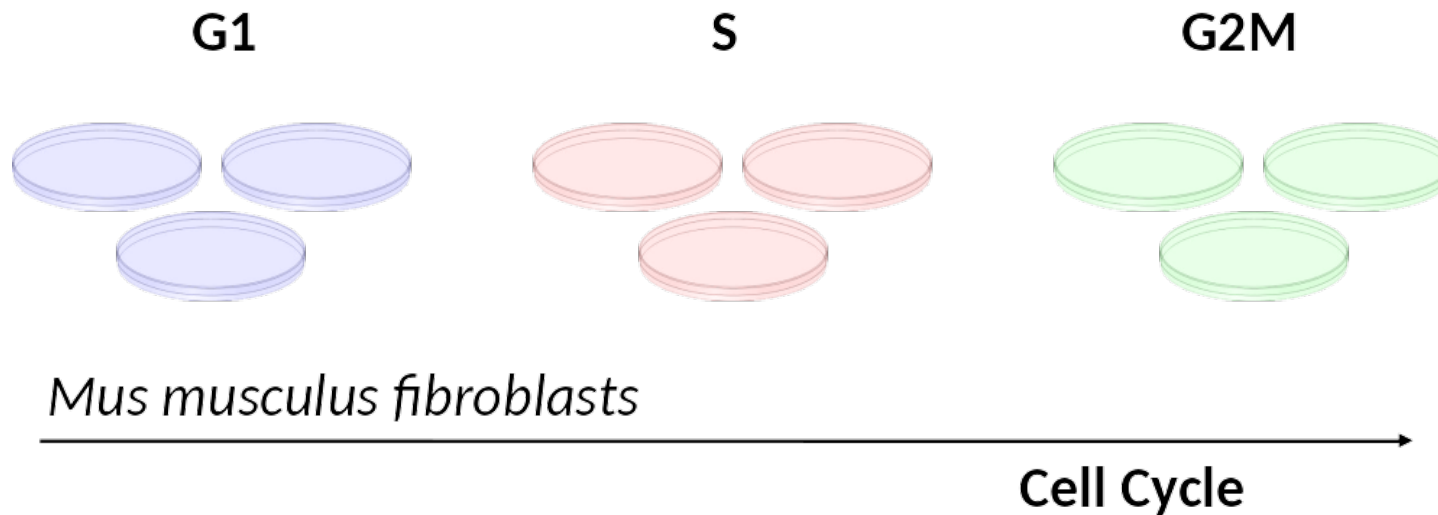




## Hands-on tutorial

*PRM method development and step-by-step analysis of Lumos PRM data*

**31 peptides from 19 proteins related to the cell cycle of mouse fibroblasts**





## Hands-on tutorial

### *PRM method development and step-by-step analysis of Lumos PRM data*

1. Setting up a Skyline document for PRM acquisition
2. Prepare and export the PRM method
3. PRM data analysis



## Hands-on tutorial

*PRM method development and step-by-step analysis of Lumos PRM data*



**EMBO Practical Course on Targeted Proteomics**  
Barcelona, 11-16 November 2018

*Registration Opens March 2018*

### Other courses

<https://skyline.ms/project/home/software/Skyline/events/begin.view?>

# Learn More

- Webinar #18: TBD (coming soon)
- Weeklong Courses 2018
  - Buck Institute, Novato, CA – April 2-6
  - NEU, Boston – April 30 – May 11
  - ETH, Zurich – July 2-6
  - U. of Wa., Seattle – July 30 – August 3
  - **New!** Duke, Durham, NC – September 17-21
  - **CRG, Barcelona – November 12-16**
  - Shanghai – October 22-26
- Workshops and Conferences 2018
  - MSACL, Palm Springs – January 20&21
  - pre-Lorne, Melbourne – January 29-31
  - IIT Bombay, Mumbai – February
  - US HUPO, Minneapolis – March 10&11
  - ASMS, San Diego – June 2&3
  - *User Group Meeting at ASMS, San Diego – June 3*
  - CNPEM, Campinas, Brazil – November 7-9

Listings updated in **Join Us** section of Skyline homepage:

<https://skyline.ms/Skyline.url>

# Questions?

- Ask any questions at the following form:

<https://skyline.ms/QA4Skyline.url>

- Take the post-webinar survey:

<https://skyline.ms/survey4webinar.url>

The logo features a stylized blue skyline with a prominent tower on the left and several rectangular buildings of varying heights to its right, all set against a white background.

# Skyline Tutorial Webinar #17

This ends this Skyline Tutorial Webinar.

Please give us feedback on the webinar at the following survey:

<https://skyline.ms/survey4webinar.url>

A recording of today's meeting will be available shortly at the Skyline website.

We look forward to seeing you at a future Skyline Tutorial Webinar.