

The Mtb Proteome Library: Development and application of assays for targeted MS analysis of the complete proteome of *Mycobacterium tuberculosis* by SRM and SWATH-MS



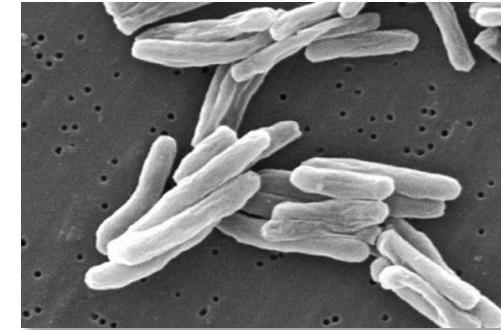
Olga Schubert

Group of Prof. Ruedi Aebersold
ETH Zurich

Skyline User Meeting 2013 Minneapolis

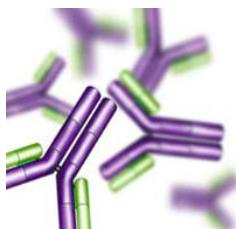


Mycobacterium tuberculosis (Mtb)



- Mycobacterium tuberculosis is the causative agent of Tuberculosis (TB)
- One third of the world's population latently infected with Mtb
- 1.7 million deaths from TB each year
- More efficient treatments urgently needed
- **Limited availability of techniques to measure proteins with high sensitivity, selectivity and reproducibility**

Traditional approach: Antibodies

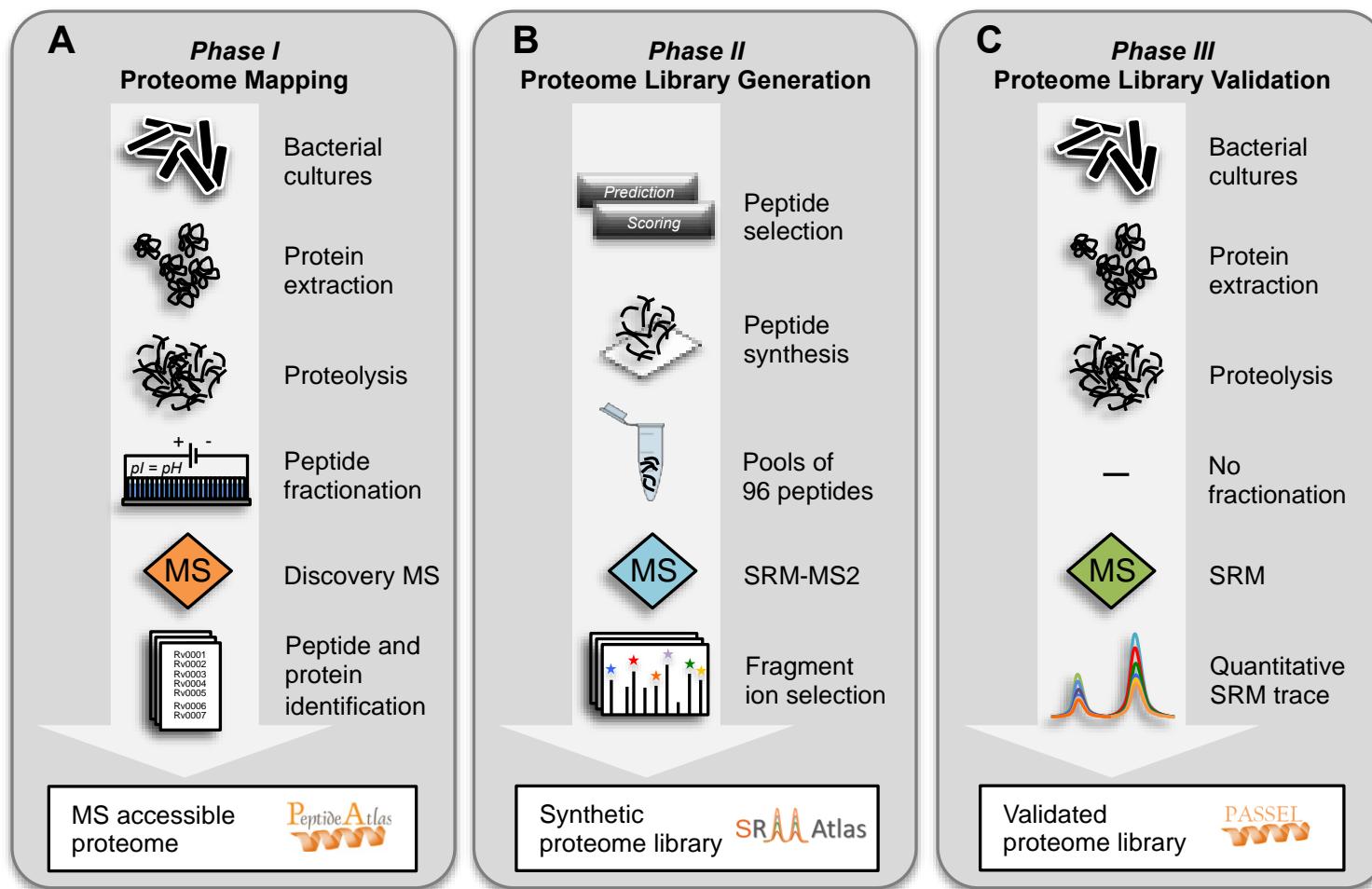


Only for few targets,
high cost, low
throughput

Aim

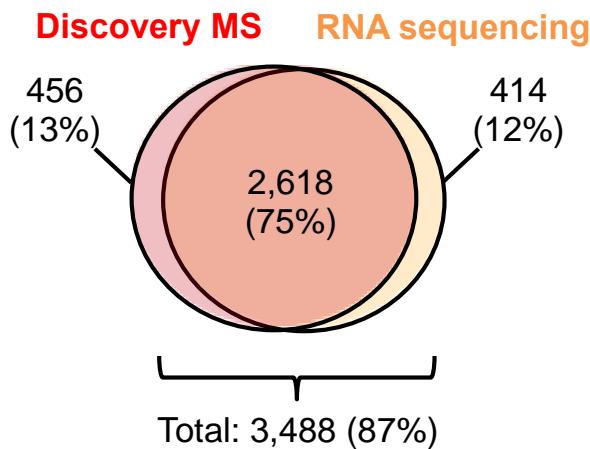
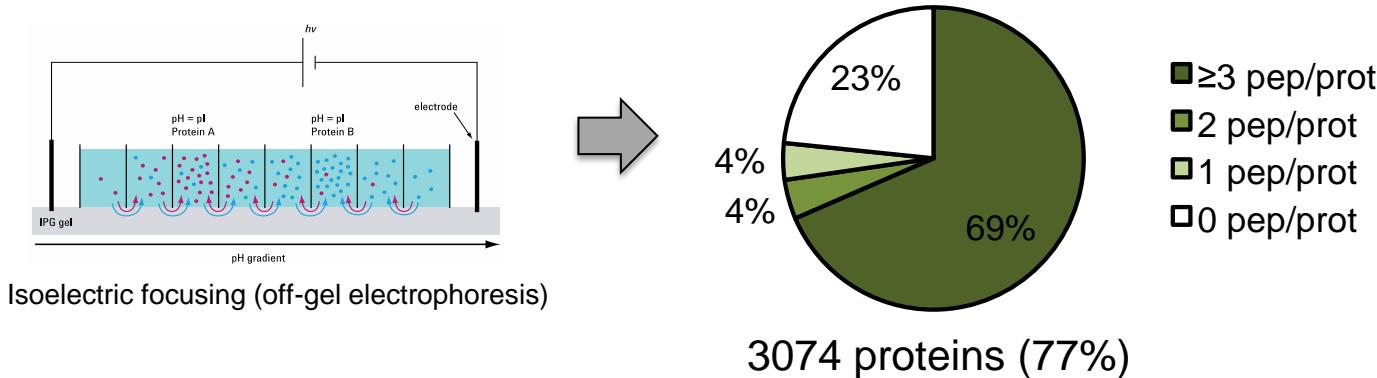
To generate a resource of validated assays for the sensitive detection and accurate quantification of every protein of *Mycobacterium tuberculosis*, even in complex backgrounds.

The Mtb Proteome Library contains SRM assays for the entire proteome of Mtb



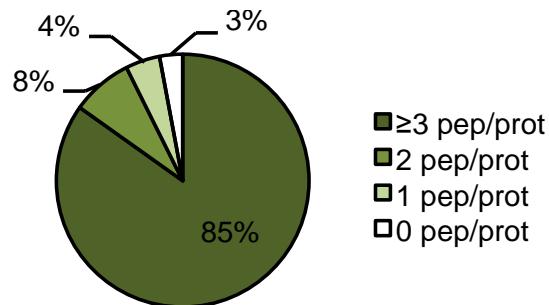
Definition of the MS-accessible Mtb proteome by discovery MS

Extensive fractionation and shotgun MS on Orbitrap XL

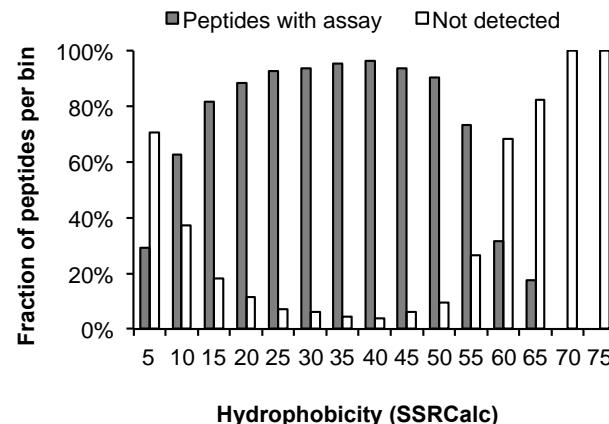
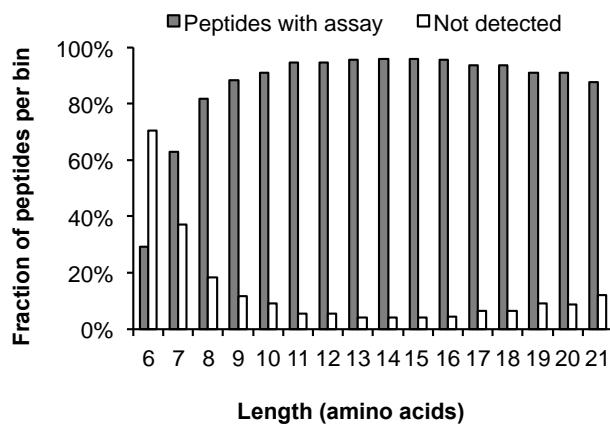
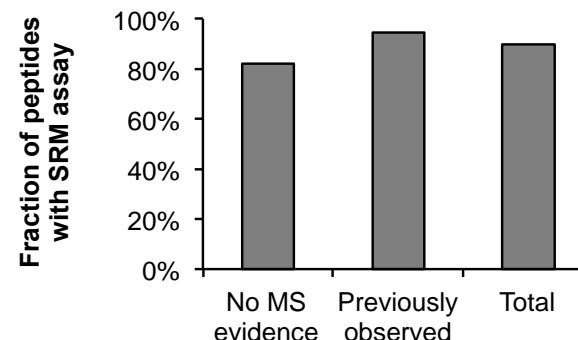


100% corresponds to the 4,012 annotated ORFs in Mtb (TubercuList v2.3)

Generation of SRM assays using crude synthetic peptides

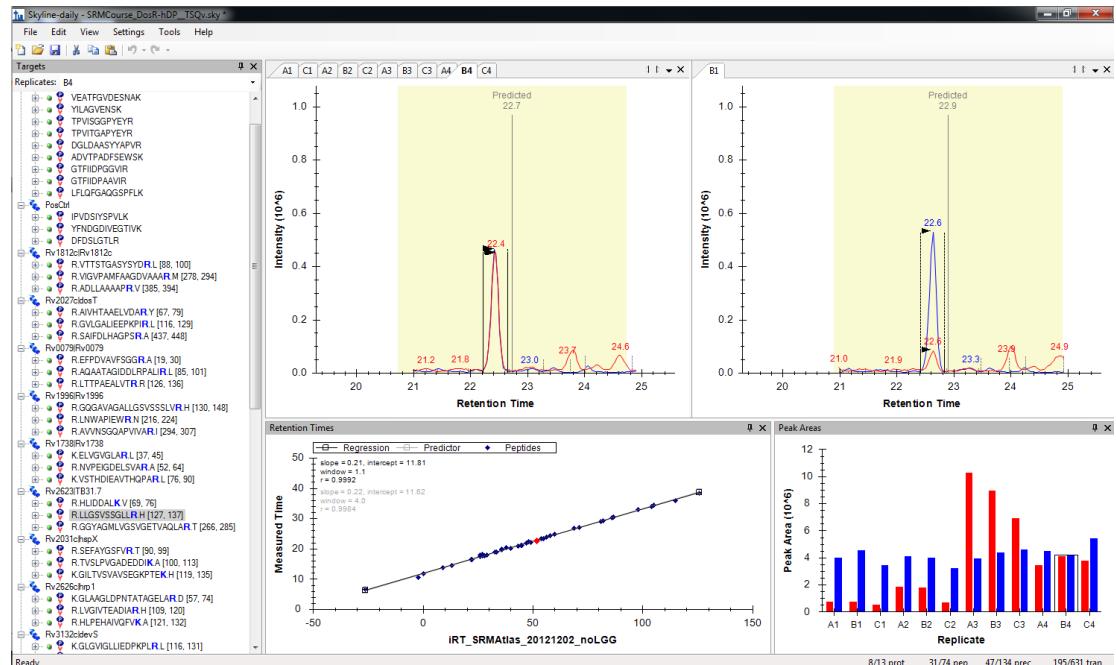
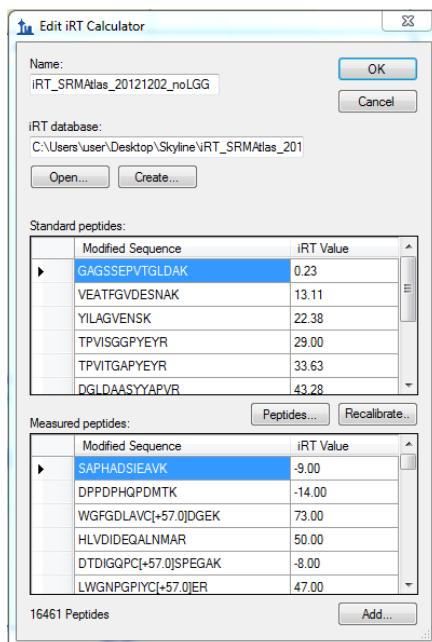
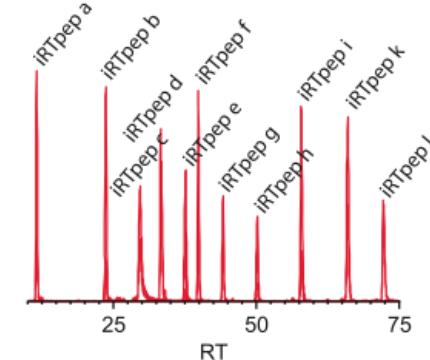


3894 proteins (97%)



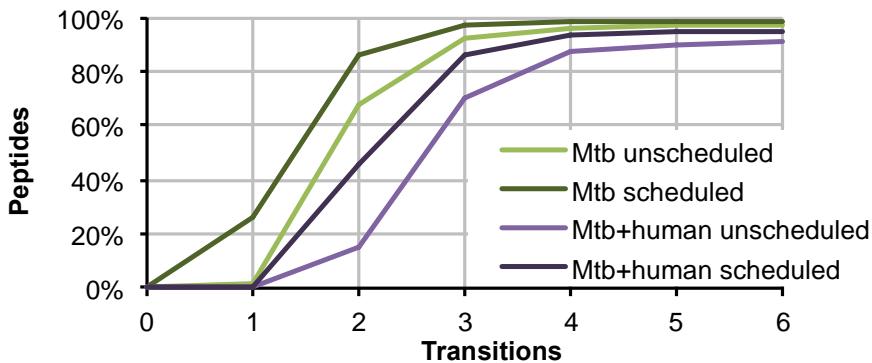
17,463 crude synthetic peptides (JPT) measured in pools of ~100 on a Qtrap 4000 in SRM-triggered MS2 mode

Increasing SRM/SWATH assay specificity and throughput by using iRTs and scheduled SRM



iRT peptides spiked into each sample allow to determine a chromatography-independent retention time (iRT) for each peptide.

Theoretical assessment of SRM assay specificity using the SRMCollider



SRM Collider

version 1.4
Hannes Röst 2012

Collider Download About Instructions

The SRMCollider is a program that will take your input transitions and compare them to all other transitions in a given background proteome and find interferences. It will report these interferences on a per-peptide basis, allowing a researcher to identify peptides that share many transitions with the target peptide.

Please enter the peptide sequences here (see [Instructions](#) for help):

```
LSDVVDLQDVQEIR[166]/2, LSDVVDLQDVQEIR[166], 2, 929.987772, 873.44574, #N/A, 2, y15
LPDNGIELC[160]R/2, LPDNGIELC[160]R, 2, 612.308612, 1031.473119, #N/A, 1, y9
LPDNGIELC[160]R/2, LPDNGIELC[160]R, 2, 612.308612, 915.446176, #N/A, 1, y8
LPDNGIELC[160]R/2, LPDNGIELC[160]R, 2, 612.308612, 582.276257, #N/A, 1, y4
LPDNGIELC[160]R/2, LPDNGIELC[160]R, 2, 612.308612, 453.233664, #N/A, 1, y3
LPDNGIELC[160]R/2, LPDNGIELC[160]R, 2, 612.308612, 561.76658, #N/A, 2, y10
LPDNGIELC[160]R[166]/2, LPDNGIELC[160]R[166], 2, 617.312746, 1041.481388, #N/A, 1, y9
LPDNGIELC[160]R[166]/2, LPDNGIELC[160]R[166], 2, 617.312746, 925.454445, #N/A, 1, y8
LPDNGIELC[160]R[166]/2, LPDNGIELC[160]R[166], 2, 617.312746, 592.284526, #N/A, 1, y4
LPDNGIELC[160]R[166]/2, LPDNGIELC[160]R[166], 2, 617.312746, 463.241933, #N/A, 1, y3
LPDNGIELC[160]R[166]/2, LPDNGIELC[160]R[166], 2, 617.312746, 566.770714, #N/A, 2, y10
```

SSRCalc window: 10 arbitrary units

Q1 mass window: 0.7 Th

Q3 mass window: 1.0 Th

Low mass threshold for transitions: 300 Th

High mass threshold for transitions: 1500 Th

Genome: Mycobacterium bovis

Consider isotopes up to: 3 amu

Missed Cleavages: 1

Find UIS up to order*: 2

Skyline-daily - SRMCourse_DosR-hDP_TSQv.sky *

File Edit View Settings Tools Help

SRM Collider

Targets

Replicates: A1

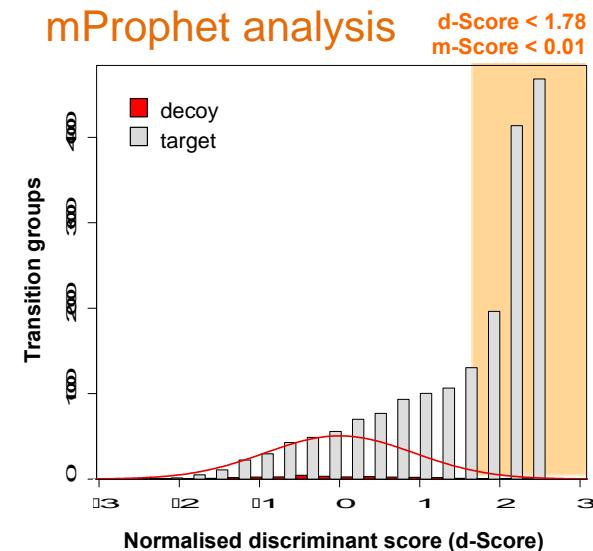
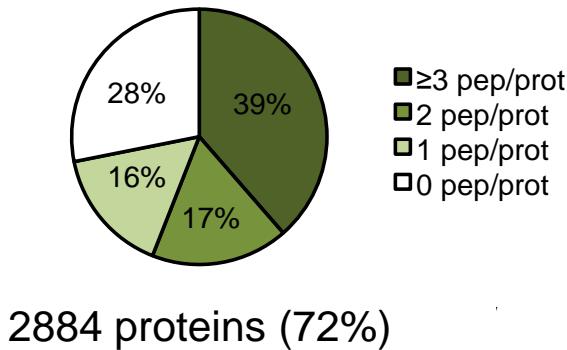
iRTpeptides

- LGGNNEQVTR
- GAGSSEPVTLDAK
- VEATFGVDESNAK
- YILAGVENSK
- TPVISGGPYEYR
- TPVITGAPYEYR
- DGLDAASYYAPVR
- ADVTRADFSEWSK

A1

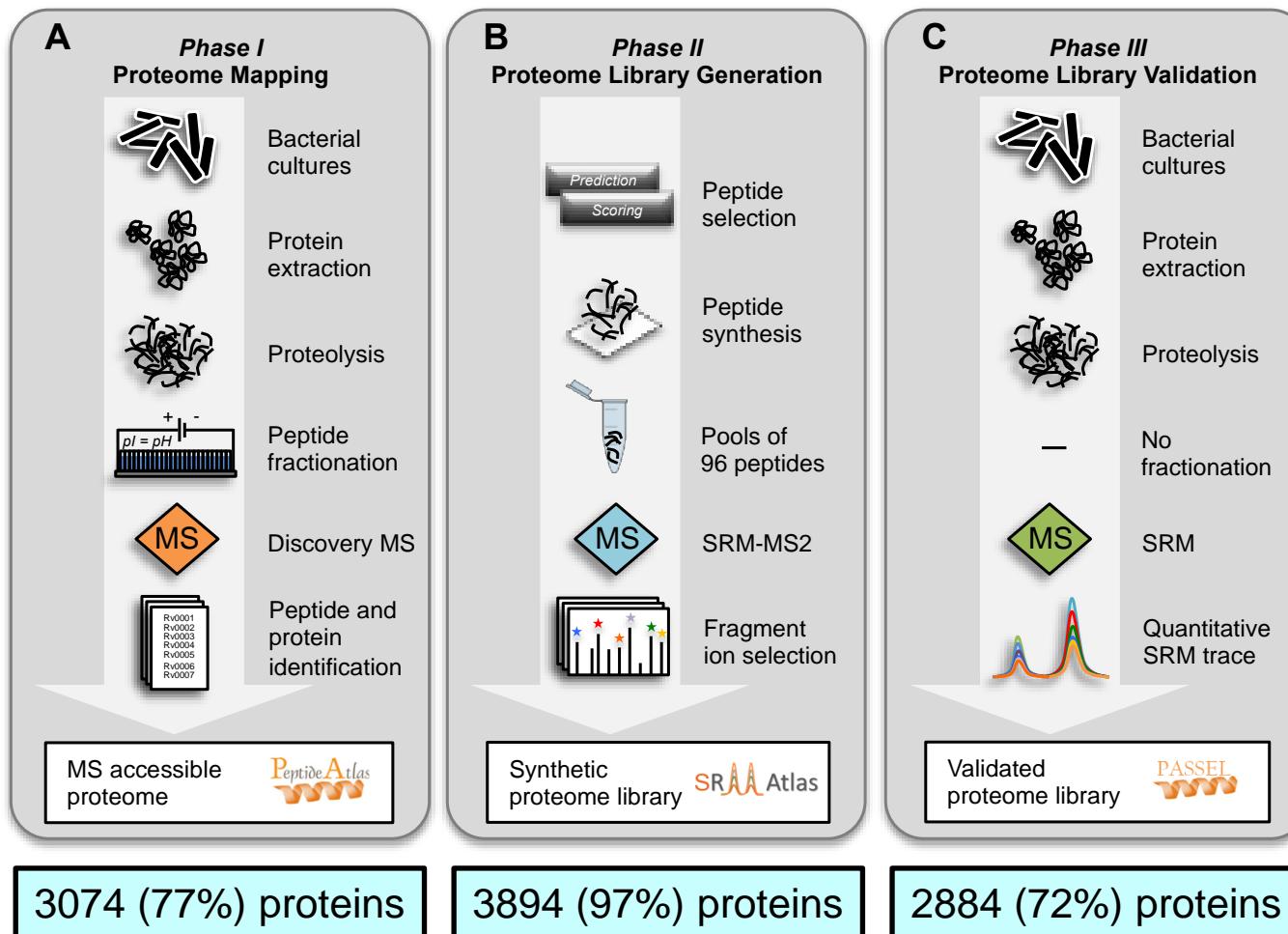
Intensity (10⁶)

Validation of the Mtb Proteome Library in unfractionated whole cell lysates by SRM



To validate all these SRM assays, over 200 scheduled SRM runs were needed.

The Mtb Proteome Library contains SRM assays for the entire proteome of Mtb



The Mtb Proteome Library is a publicly available resource of MS reference data, SRM assays and their validation

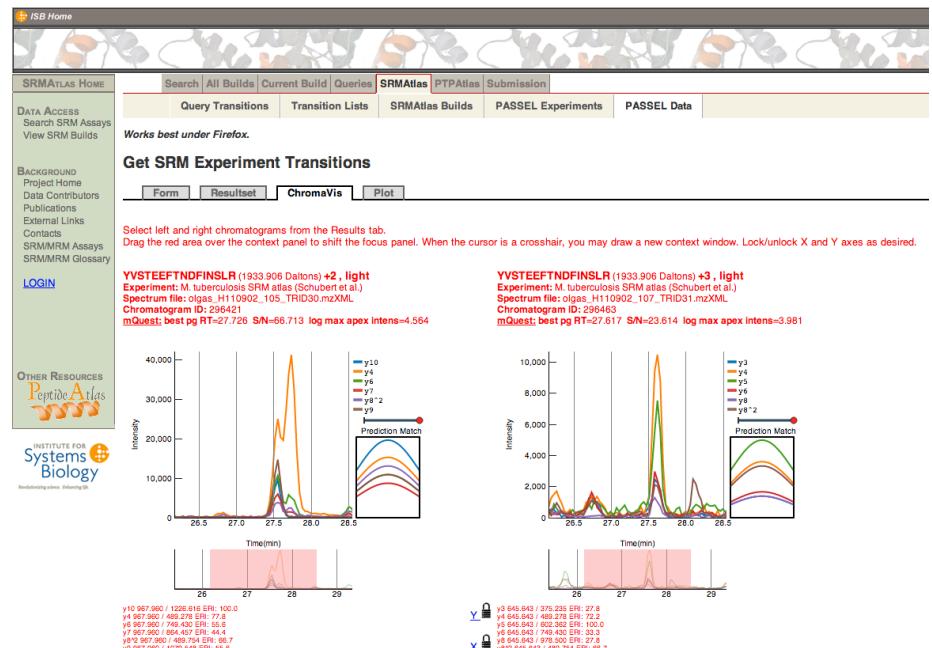
www.PeptideAtlas.org

This screenshot shows the SRMAtlas Home page with a search bar for "Rv0001". Below the search bar, a protein summary for Rv0001 is displayed, including its name, gene name (dnaA), description (catalase), and various statistics like distinct peptides (31) and total observations (443). A sequence motif plot is shown below the summary, followed by sequence position and coverage plots. A sequence display mode section for Trypsin is also present.

www.SRMAtlas.org

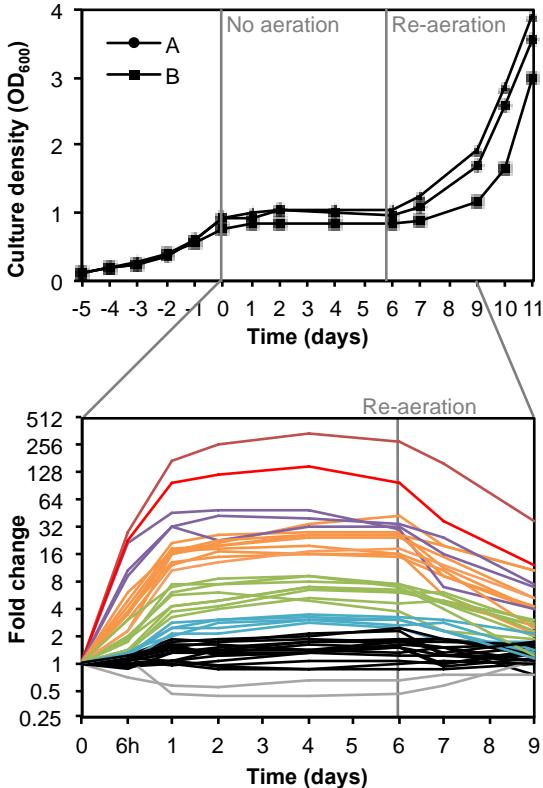
This screenshot shows a table of peptide sequences from the SRMAtlas Home page. The columns include Protein, Pre AA, Sequence, For AA, Adj SS, Source, Q1_ms, Q1_chg, Q3_ms, Q3_chg, Ion, Name, RI, SIRRT, N_map, and IonTrap. The table lists several peptides, such as Rv0001, VYSTEETNDFINSLR, and FAHAAALAAIAEAPAR, with their respective properties and ion trap fragmentation patterns.

www.PeptideAtlas.org/PASSEL



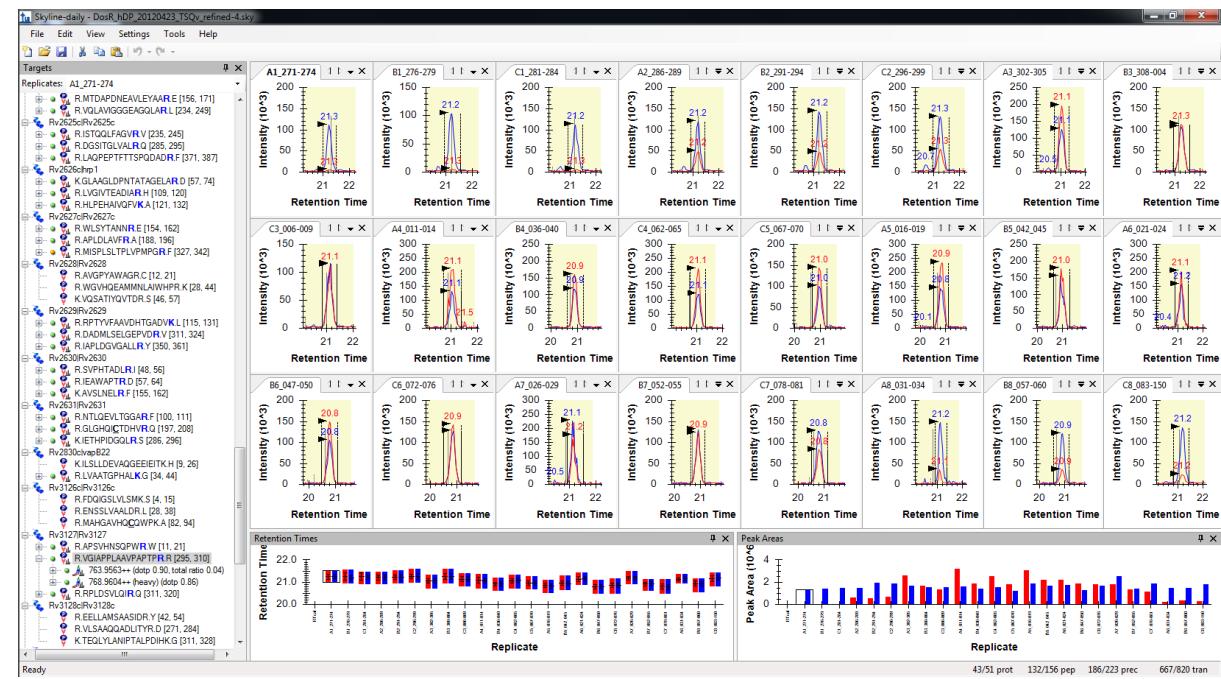
Application of the Mtb Proteome Library to study the dynamics of the DosR regulon of Mtb under hypoxia

Mycobacterium bovis BCG

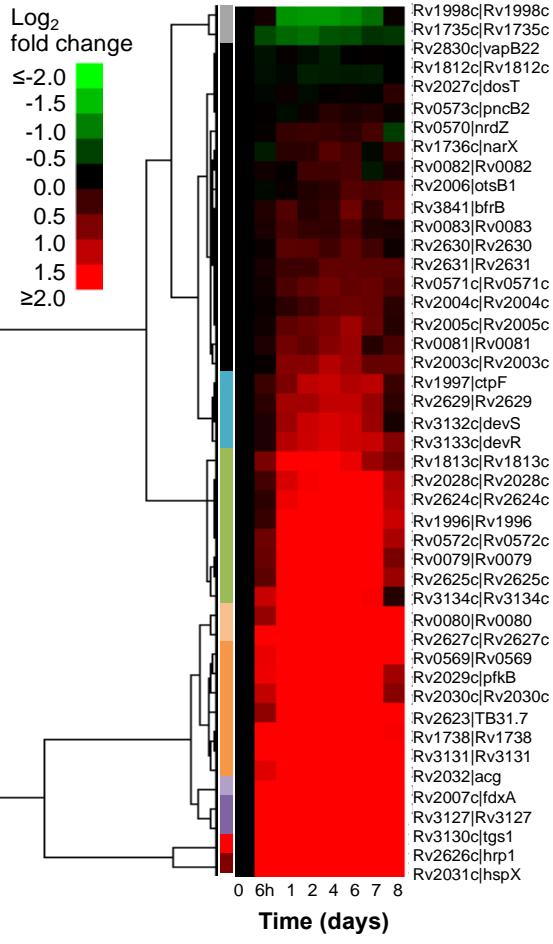


DosR study summary

Total number of proteins	53
Proteins with SRM assays	52
Proteins with validated assays in exponential and stationary growth phase	37
Proteins detected in DosR study	45

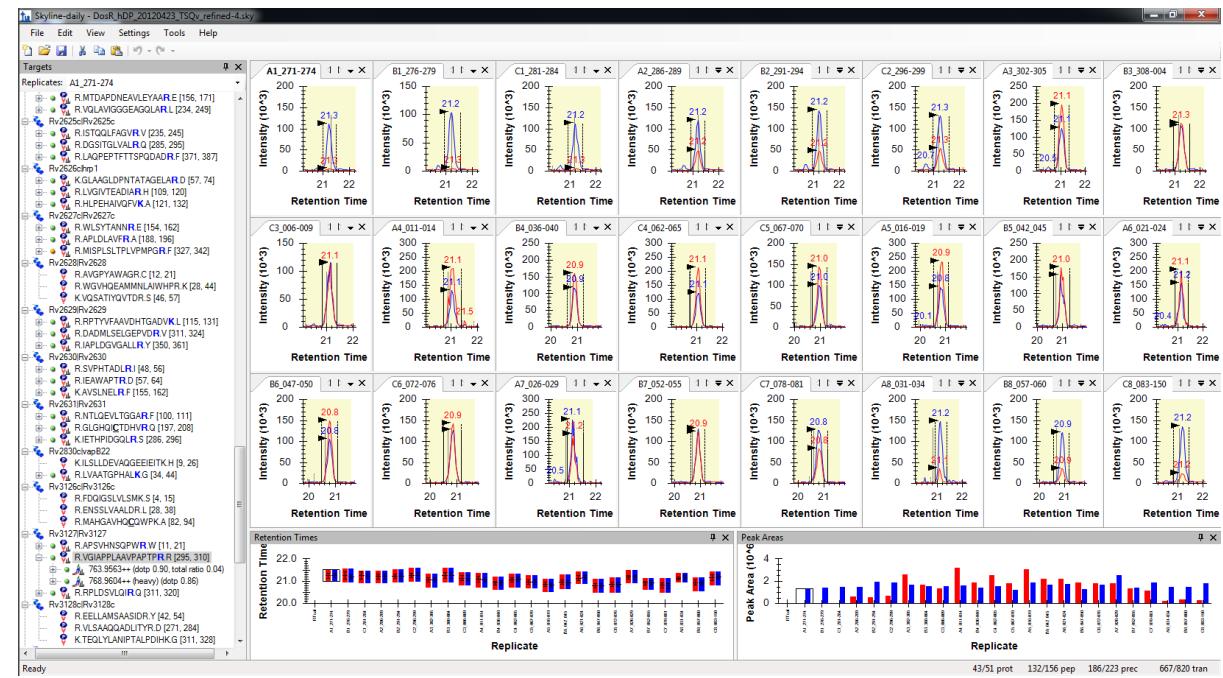


Application of the Mtb Proteome Library to study the dynamics of the DosR regulon of Mtb under hypoxia



DosR study summary

Total number of proteins	53
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Statistical analysis of SRM data by SRMstats

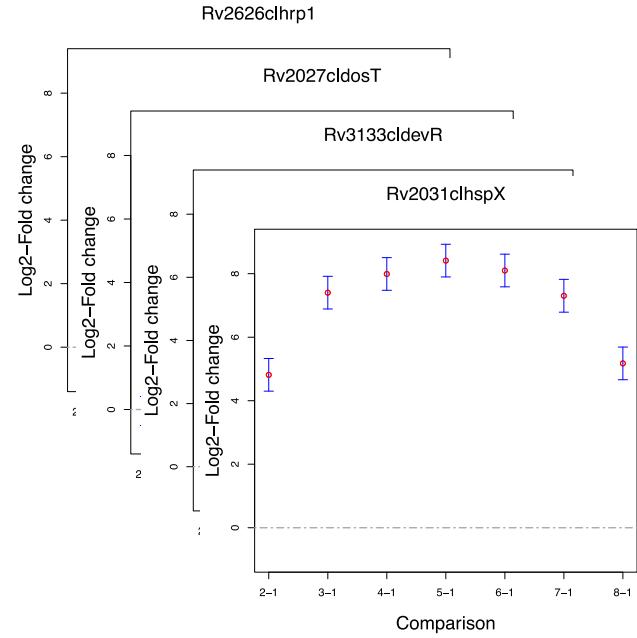
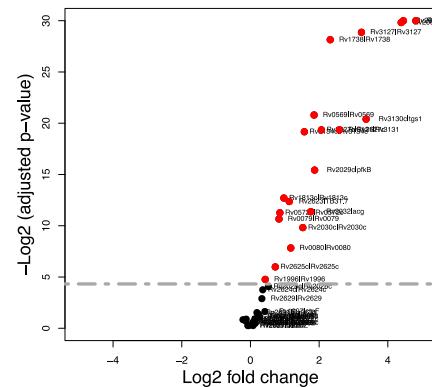
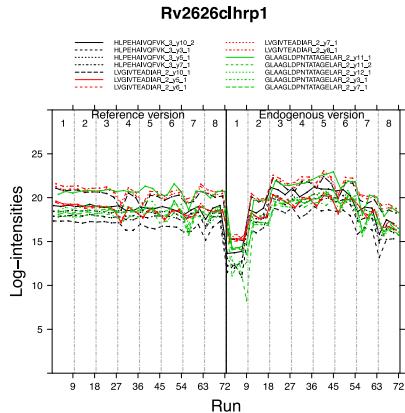


SRMstats

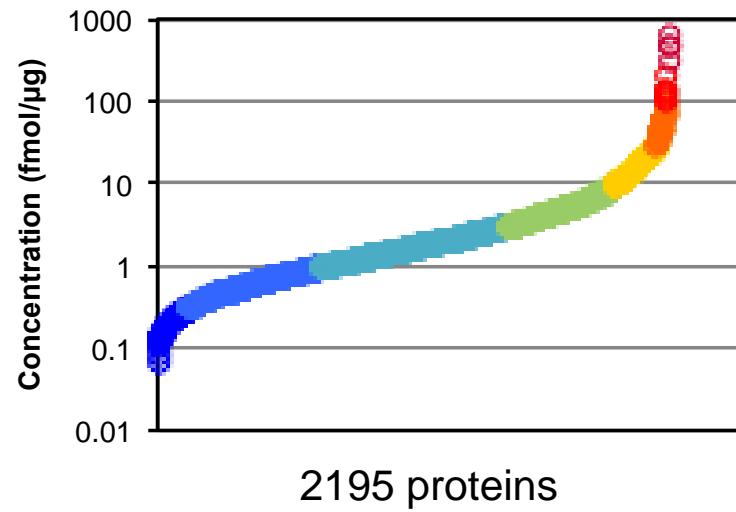
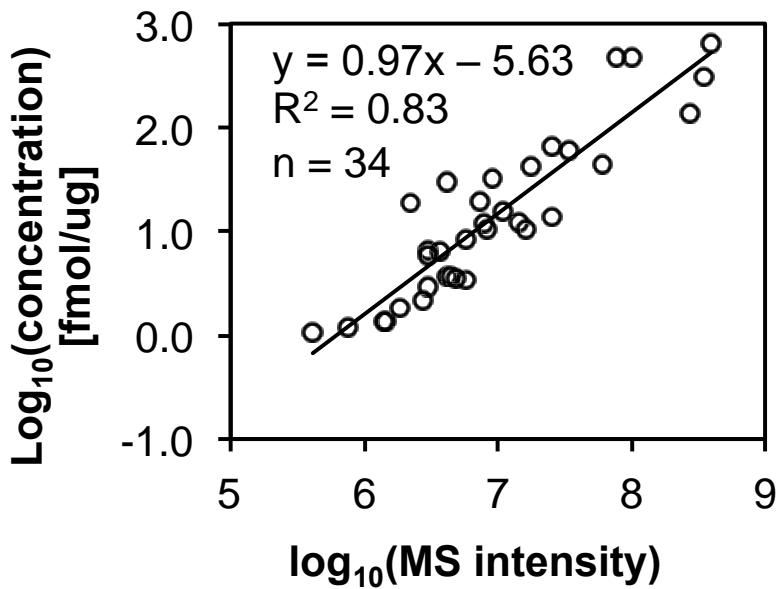
Protein significance analysis in SRM measurements

Goal

A statistical framework is proposed for protein quantification in SRM experiments based on a family of linear mixed-effects models. The framework is sensitive and flexible, and is applicable to a variety of experimental designs and to both label-based and label-free workflows.

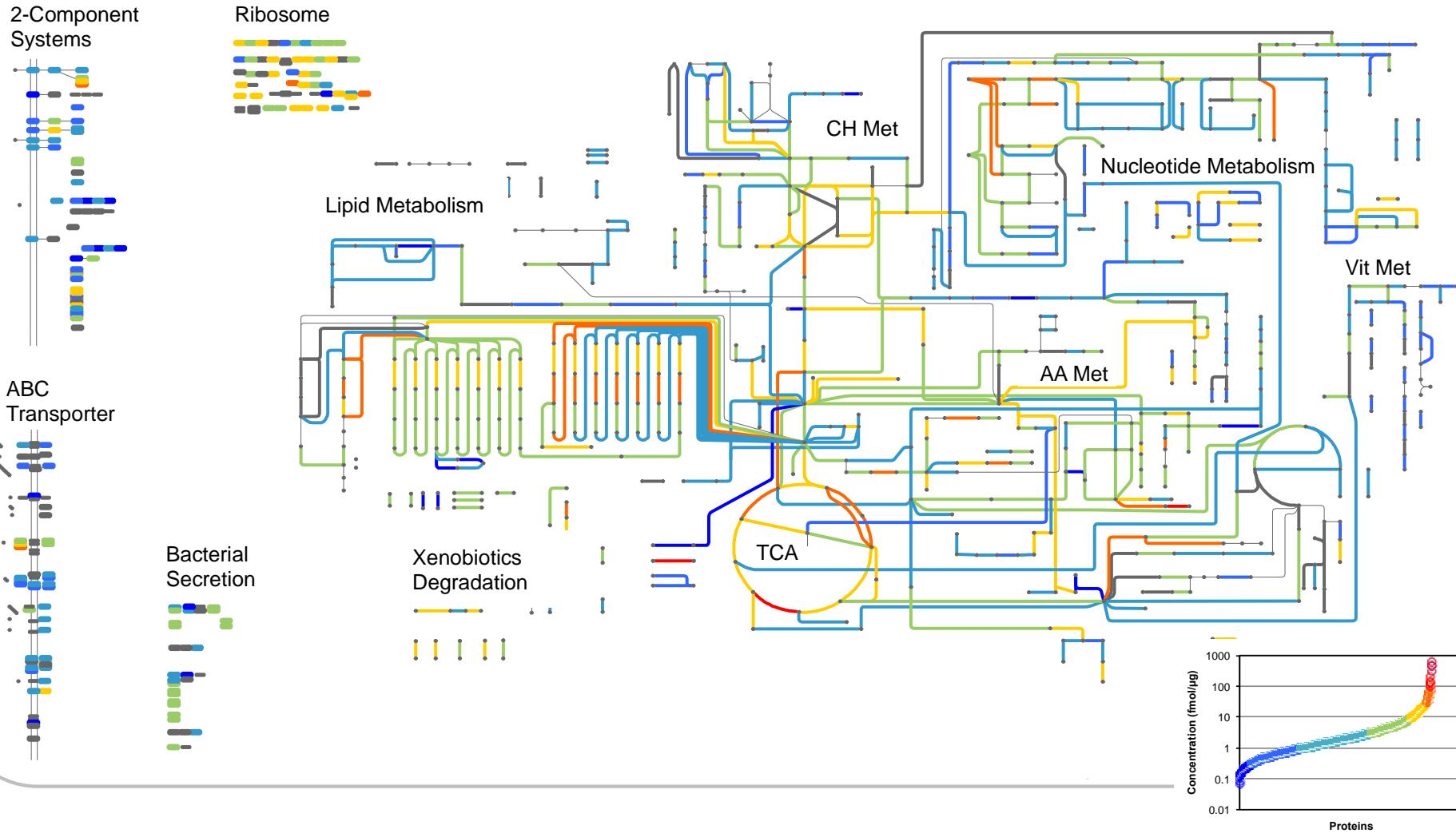


Absolute label-free quantification by SRM exploits the linear correlation of the sum of the top transitions of the top peptides per protein and the protein concentration

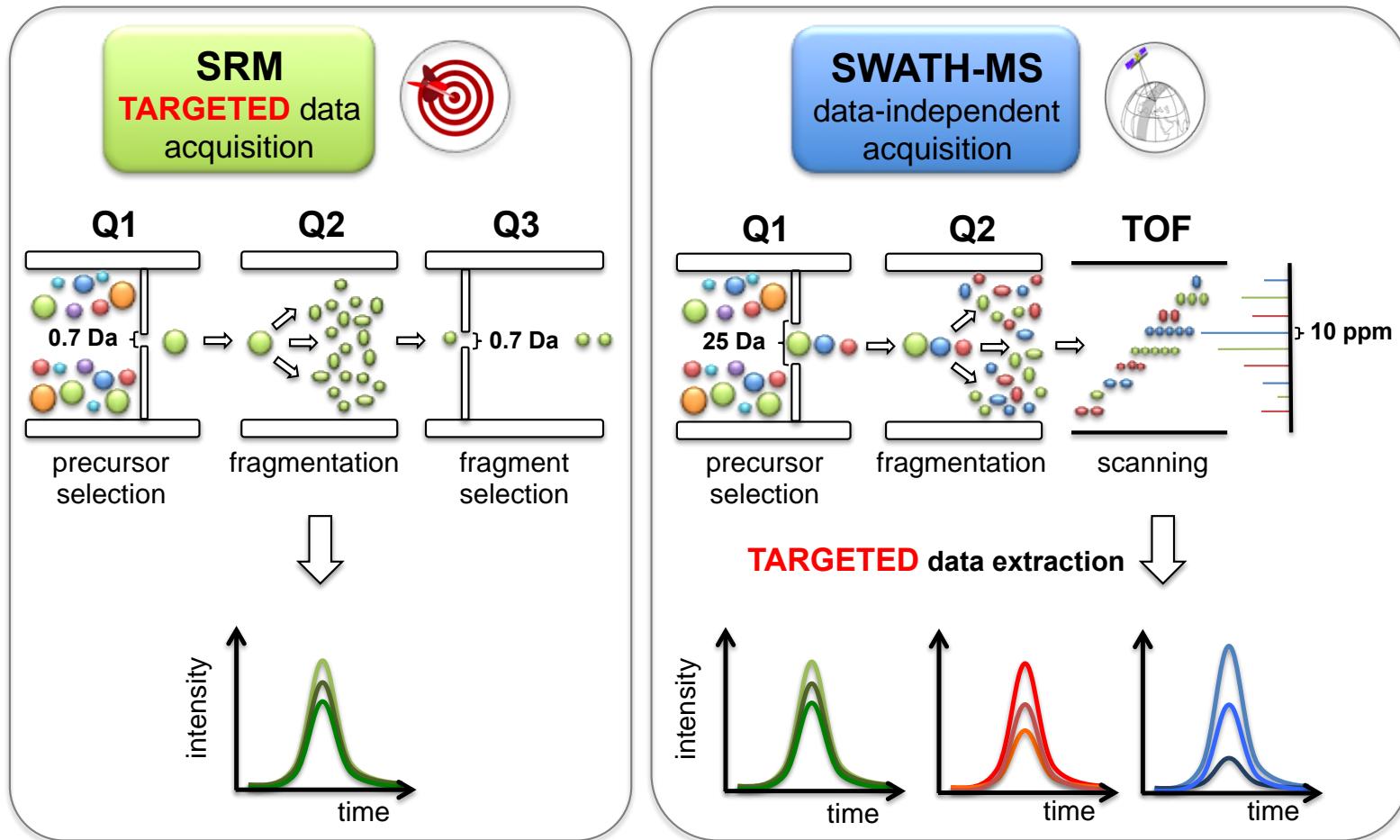


Linear correlation established using 34 anchor proteins quantified by AQUA peptides
MS intensity: sum of 2 most intense transitions of the 3 most intense peptides per protein

Proteome-wide absolute abundance estimates for Mtb



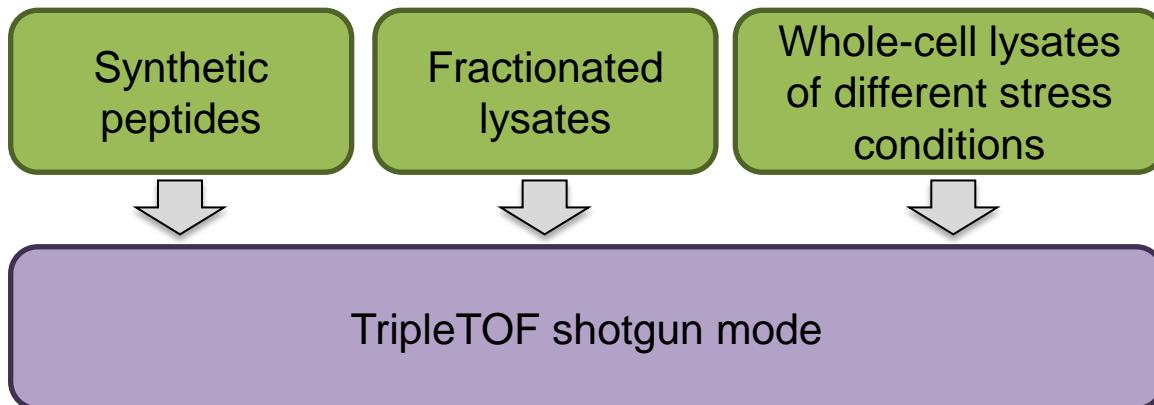
SWATH-MS: Data-independent acquisition with targeted data extraction



Gillet et al., MCP 2012, Targeted data extraction of the MS/MS spectra generated by data-independent acquisition: a new concept for consistent and accurate proteome analysis.

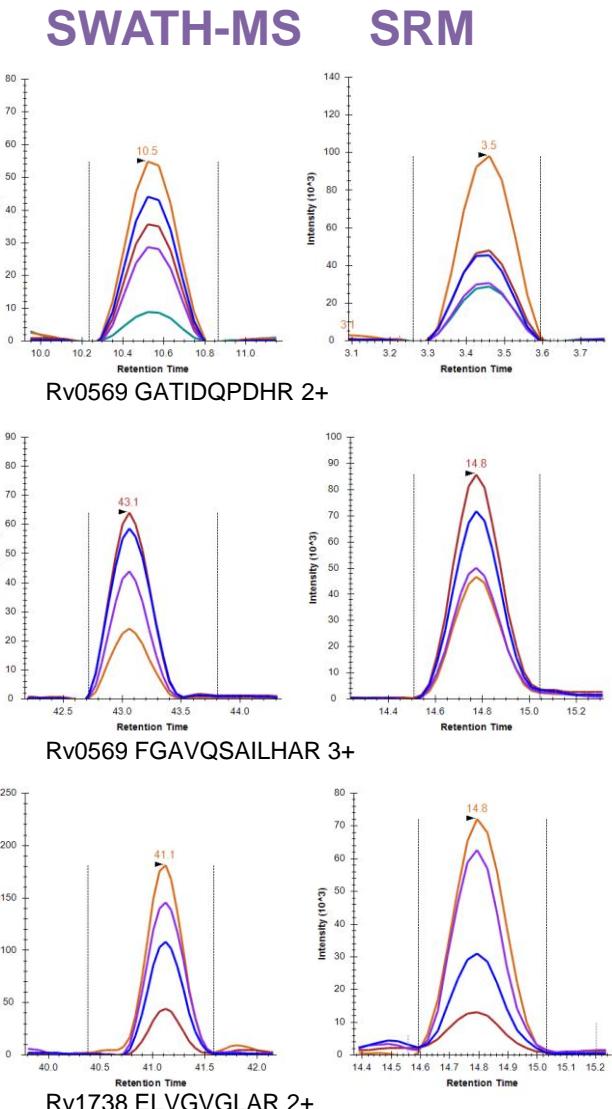
Figure by Christina Ludwig

Generation of the Mtb SWATH library

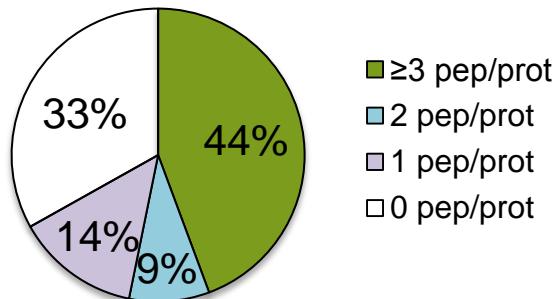


- Search each dataset with Mascot and Sequest
 - iProphet combination of all datasets
- Alignment of runs into a common RT space (iRT)
- Consensus spectral library generation with SpectraST
 - Extraction of the top transitions per precursor

Mtb SWATH library
3931 proteins (98%)
39,479 peptides

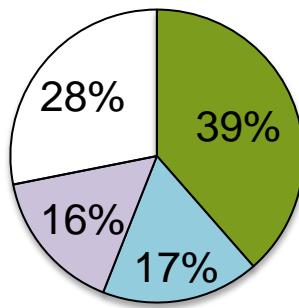


SWATH-MS allows reproducible, high proteome coverage measurements of Mtb in a single run



Proteome coverage by
SWATH-MS

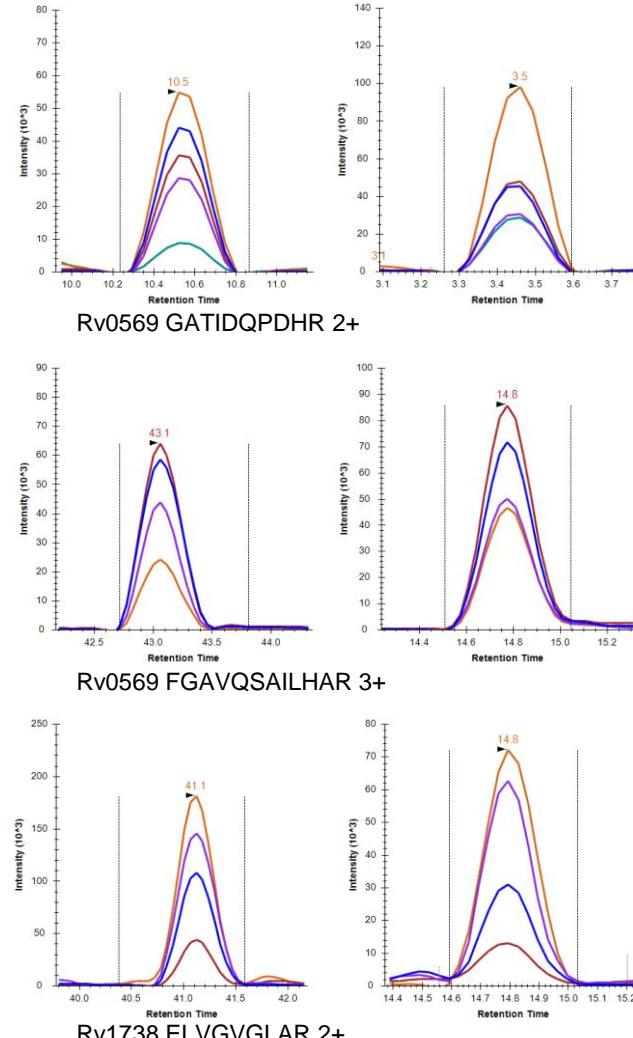
single injection
2683 proteins (67%)
(openSWATH software)



Proteome coverage by
SRM

>200 injections
2884 proteins (72%)
(mProphet software)

SWATH-MS SRM





Summary

- The Mtb Proteome Library is a public resource of SRM assays for the entire proteome of Mtb
 - SRM assays generated from crude synthetic peptides and validated in whole cell lysates
 - Data analysis with **Skyline** and supporting tools: iRT peptides, SRMCollider, mProphet, SRMstats
 - Data can be browsed on and downloaded from www.SRMAtlas.org, www.PeptideAtlas.org/passel
- Application of the Mtb Proteome Library to study the dynamics of the DosR regulon of Mtb under hypoxia
- Absolute label-free quantification by SRM exploits the linear correlation of the sum of the top transitions of the top peptides per protein and its absolute concentration.
- Expansion of the Mtb Proteome Library for use with SWATH-MS
- SWATH-MS allows reproducible, high proteome coverage measurements of Mtb in a single run

ETH Zurich

- Prof. Ruedi Aebersold
- Christina Ludwig
- Jeppe Mouritsen
- George Rosenberger
- **Hannes Röst (Mon 5:45 pm and WP 595, Wed)**
- **Ludovic Gillet (TOD pm, Tue 5:30 pm)**
- **Ben Collins (WP 688, Wed)**
- Alessio Maiolica, Mariette Matondo

University of Ghana

- Dr. Patrick K. Arthur

Max Planck Institute Berlin

- Prof. Stefan Kaufmann
- Dr. Martin Gengenbacher

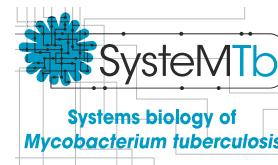
ISB Seattle

- Prof. Rob Moritz
- Dave Campbell
- Zhi Sun
- Terry Farrah
- **Samuel Bader (ABSciex, Mon 7 am)**

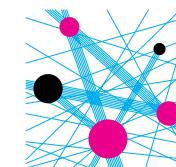
University of Washington

- **Brendan MacLean (TP 499, Tue)**

Acknowledgements



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Institute of
Molecular
Systems
Biology